

Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.2.2 Annex Point/TNsG Annex IIIA, XII.1.1, Annex VI, para 85	Field soil dissipation and accumulation Section 7: Ecotoxicological Profile, including Fate and Behaviour	
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only
Other existing data <input type="checkbox"/>	Technically not feasible <input type="checkbox"/>	Scientifically unjustified <input checked="" type="checkbox"/>
Detailed justification:	<p>Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required.</p> <p>Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.</p>	
Undertaking of intended data submission <input type="checkbox"/>	Not applicable.	



Section 7.2.2.2 Annex Point/TNsG Annex IIIA, XII.1.1, Annex VI, para 85	Field soil dissipation and accumulation Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
	Use separate “evaluation boxes” to provide transparency as to the comments and views submitted
EVALUATION BY RAPPORTEUR MEMBER STATE Date Evaluation of applicant’s justification Conclusion Remarks	<i>Give date of action</i> <i>Discuss applicant’s justification and, if applicable, deviating view</i> <i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
COMMENTS FROM OTHER MEMBER STATES (specify) Date Evaluation of applicant’s justification Conclusion Remarks	<i>Give date of comments submitted</i> <i>Discuss if deviating from view of rapporteur member state</i> <i>Discuss if deviating from view of rapporteur member state</i>

Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.2.3 Annex Point/TNsG Annex IIIA, XII.1.4	Extent and nature of bound residues Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input type="checkbox"/>	Technically not feasible <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Scientifically unjustified <input checked="" type="checkbox"/>
Detailed justification:	Other justification <input type="checkbox"/> Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required. Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.	
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.

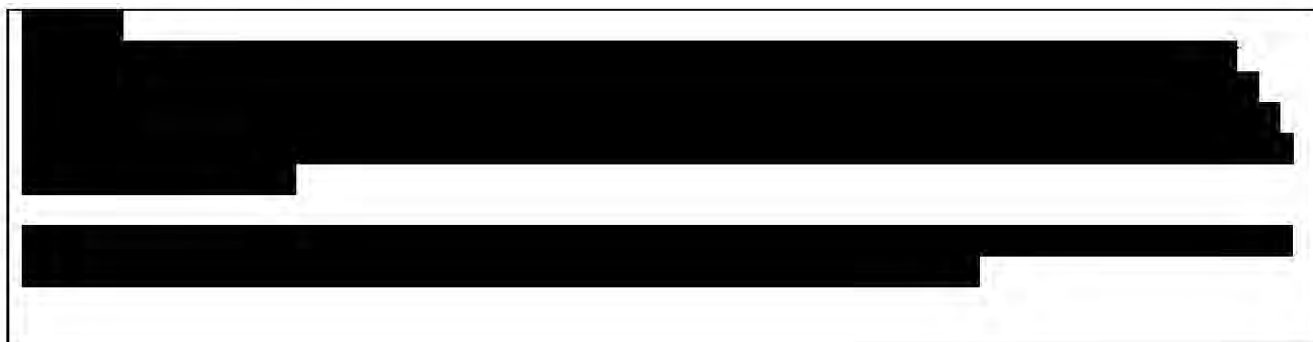


Section 7.2.2.3 Annex Point/TNsG Annex IIIA, XII.1.4	Extent and nature of bound residues Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
	Use separate “evaluation boxes” to provide transparency as to the comments and views submitted
EVALUATION BY RAPPORTEUR MEMBER STATE	
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Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.2.4 Annex Point/TNsG Annex IIIA, XII.1.1	Other soil degradation studies Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input type="checkbox"/>	Technically not feasible <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Scientifically unjustified <input checked="" type="checkbox"/>
Detailed justification:	Other justification <input type="checkbox"/> Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required. Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.	
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.



Section 7.2.2.4 Annex Point/TNsG Annex IIIA, XII.1.1	Other soil degradation studies Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.3 Annex Point/TNsG Annex IIIA, XII.1.2-1.3	Adsorption and mobility in soil, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
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Limited exposure <input checked="" type="checkbox"/>	Other justification <input type="checkbox"/>	
Detailed justification:	Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required. Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.	
Undertaking of intended data submission <input type="checkbox"/>	Not applicable.	

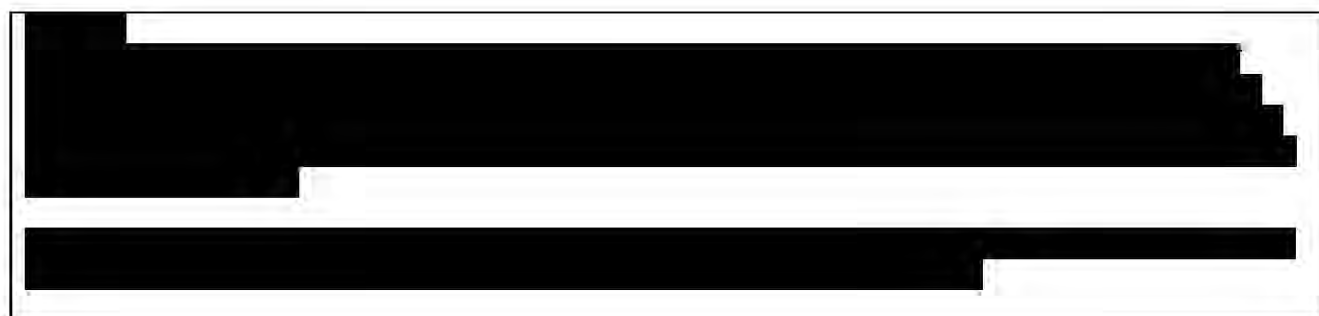


Section 7.2.3 Annex Point/TNsG Annex IIIA, XII.1.2-1.3	Adsorption and mobility in soil, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.3.1 Annex Point/TNsG Annex IIIA, XII.1.2	Adsorption and desorption in accordance with the new test guideline EC C18 or the corresponding OECD 106 and, where relevant, adsorption and desorption metabolites and degradation products Section 7: Ecotoxicological Profile, including Fate and Behaviour	
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Detailed justification:	<p>Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required.</p> <p>Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.</p>	
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.



Section 7.2.3.1 Annex Point/TNsG Annex IIIA, XII.1.2	Adsorption and desorption in accordance with the new test guideline EC C18 or the corresponding OECD 106 and, where relevant, adsorption and desorption metabolites and degradation products Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Table 4-2: Standard form for justification of the non-submission of data

Section 7.2.3.2 Annex Point/TNsG Annex IIIA, XII.1.3	Mobility in at least three soil types and where relevant mobility of metabolites and degradation products Section 7: Ecotoxicological Profile, including Fate and Behaviour	
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only
Other existing data <input type="checkbox"/>	Technically not feasible <input type="checkbox"/>	Scientifically unjustified <input checked="" type="checkbox"/>
Limited exposure <input checked="" type="checkbox"/>	Other justification <input type="checkbox"/> <p>Detailed justification:</p> <p>Carbon dioxide, as a biocide, as used by Rentokil Initial is not applied or emitted directly to the soil and therefore this study is not required.</p> <p>Data requirements A7.1.1.2.1 and A7.1.1.2.2 do not indicate the need to conduct studies on the fate and behaviour of carbon dioxide in soil and in addition, this is substantiated by the fact that carbon dioxide does undergo a degree of abiotic degradation by means of simple dissolution in water. Also, it is well known that although carbon dioxide occurs predominantly in air, it will attain equilibrium with air spaces in soil through passive diffusion.</p>	
Undertaking of intended data submission <input type="checkbox"/>	Not applicable.	



Section 7.2.3.2 Annex Point/TNsG Annex IIIA, XII.1.3	Mobility in at least three soil types and where relevant mobility of metabolites and degradation products Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Table 4-2: Standard form for justification of the non-submission of data

Section 7.3.1 Annex Point/TNsG Annex IIIA, VII.5	Phototransformation in air (estimation method), including identification of breakdown products Section 7: Ecotoxicological Profile, including Fate and Behaviour	
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only
Other existing data	<input type="checkbox"/>	Technically not feasible <input type="checkbox"/>
		Scientifically unjustified <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification <input type="checkbox"/>
<p>Detailed justification:</p> <p>As a gas, under all environmental conditions that are likely to occur on earth, carbon dioxide will occur predominantly in air. Carbon dioxide occurs as a by-product of aerobic respiration. There is a natural “Carbon Cycle” whereby carbon dioxide is continuously added and removed from the environment through natural processes.</p> <p>Under normal conditions of use, the carbon dioxide used in Rentokil Initial’s rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in air, outside normal atmospheric ranges. <i>Refer to next page for details of scientific calculation which supports this statement.</i></p> <p>(Continued...)</p>		

Section 7.3.1 Annex Point/TNsG Annex IIIA, VII.5	Phototransformation in air (estimation method), including identification of breakdown products Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
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Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
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Table 4-2: Standard form for justification of the non-submission of data

Section 7.3.2 Annex Point/TNsG Annex IIIA, XII.3	Fate and behaviour in air, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input type="checkbox"/>		
Detailed justification:	<p>As a gas, under all environmental conditions that are likely to occur on earth, carbon dioxide will occur predominantly in air. Carbon dioxide occurs as a by-product of aerobic respiration. There is a natural “Carbon Cycle” whereby carbon dioxide is continuously added and removed from the environment through natural processes.</p> <p>Under normal conditions of use, the carbon dioxide used in Rentokil Initial’s rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in air, outside normal atmospheric ranges.</p> <p><i>Refer to next page for details of scientific calculation which supports this statement.</i></p> <p align="center">(Continued.....)</p>				



Section 7.3.2 Annex Point/TNsG Annex IIIA, XII.3	Fate and behaviour in air, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Table 4-2: Standard form for justification of the non-submission of data

Section A7.4.1.1 Annex Point/TNsG Annex IIA, VII 7.1	Acute toxicity to fish Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input checked="" type="checkbox"/>	Technically not feasible <input type="checkbox"/> Scientifically unjustified <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification <input type="checkbox"/>
Detailed justification:	<p>An acute toxicity study for carbon dioxide to fish is not considered necessary for the following reasons:</p> <ol style="list-style-type: none"> 1. It is not scientifically necessary because under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water, outside normal atmospheric ranges. <i>See next page for full details of the scientific calculation, which supports this statement.</i> 2. Study summaries are available for two pieces of work investigating the effects of carbon dioxide to fish. One of which shows that dissolved concentrations of up to 6.3% carbon dioxide have not given rise to irreversible physiological and behavioural effects. <i>Refer to study summaries for details about the data available on the inhalation toxicity of carbon dioxide.</i> <p>Given that substantial rises in atmospheric carbon dioxide is never going to be reached under the normal conditions of use of Rentokil Initial's biocidal products and the data cited in this application, it is scientifically unjustified to conduct further studies.</p> <p>Continued....</p>	



[REDACTED]

Section A7.4.1.1 Annex Point/TNsG Annex IIA, VII 7.1	Acute toxicity to fish Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:
(continued):

[REDACTED]

[REDACTED]

[REDACTED]

Undertaking of intended data submission Not applicable

[REDACTED]

[REDACTED]

Section A7.4.1.1 Annex Point/TNsG Annex IIA, VII 7.1	Acute toxicity to fish Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Remarks	

**Section A7.4.1.1
Annex Point IIA7.1**

Acute toxicity to fish (1 of 2)

Official
use only

1 REFERENCE

1.1 Reference

[Redacted]

1.2 Data protection

[Redacted]

1.2.1 Data owner

[Redacted]

1.2.3 Criteria for data protection

[Redacted]

2 GUIDELINES AND QUALITY ASSURANCE

2.1 Guideline study

No.

Not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075.

2.2 GLP

No.

No details available regarding the testing guidelines used.

2.3 Deviations

Yes.

This study did not follow the test guidelines of Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075. Rather than looking at acute toxicity *per se*, this investigates the physiological and behavioural effects of fish exposed to carbon dioxide.

3 MATERIALS AND METHODS

3.1 Test material

As given in section 2.

3.1.1 Lot/Batch number

Not reported.

3.1.2 Specification

[Redacted]

3.1.3 Purity

[Redacted]

Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (1 of 2)

3.1.4	Composition of product	Not applicable for active substance.																																										
3.1.5	Further relevant properties	Not reported.																																										
3.1.6	Method of analysis	Not reported.																																										
3.2	Preparation of TS solution for poorly soluble or volatile test substances	Not applicable.																																										
3.3	Reference substance	No.																																										
3.3.1	Method of analysis for reference substance	Not applicable as reference substance was not used.																																										
3.4	Testing procedure																																											
3.4.1	Dilution water	<table border="1"> <thead> <tr> <th>Criteria</th> <th colspan="3">Details</th> </tr> </thead> <tbody> <tr> <td>Source</td> <td colspan="3">Not reported.</td> </tr> <tr> <td>Alkalinity</td> <td colspan="3">23.5 mg/L</td> </tr> <tr> <td>Hardness</td> <td colspan="3">35.1 mg/L</td> </tr> <tr> <td>pH</td> <td colspan="3">6.8</td> </tr> <tr> <td>Oxygen content</td> <td colspan="3">11.2 mg/L</td> </tr> <tr> <td>Conductance</td> <td colspan="3">89 µmhos</td> </tr> <tr> <td>Holding water different from dilution water</td> <td colspan="3">Laboratory holding and test waters have same properties.</td> </tr> </tbody> </table>			Criteria	Details			Source	Not reported.			Alkalinity	23.5 mg/L			Hardness	35.1 mg/L			pH	6.8			Oxygen content	11.2 mg/L			Conductance	89 µmhos			Holding water different from dilution water	Laboratory holding and test waters have same properties.										
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3.4.2	Test organisms	<table border="1"> <thead> <tr> <th>Criteria</th> <th colspan="3">Details</th> </tr> </thead> <tbody> <tr> <td>Species / strain</td> <td>Brook trout <i>Salvelinus fontinalis</i></td> <td>Slimy Sculpin <i>Cottus cognatus</i></td> <td>Blacknose dace <i>Rhinichthys atratulus</i></td> </tr> <tr> <td>Source</td> <td>Private hatchery</td> <td>Local second-order streams</td> <td>Local second-order streams</td> </tr> <tr> <td>Wild caught</td> <td>No</td> <td>Yes – no other details given</td> <td>Yes – no other details given</td> </tr> <tr> <td>Age / size</td> <td>Age not given 170 ± 11 mm total length</td> <td>Age not given 70 ± 4 mm total length</td> <td>Age not given 69 ± 4 mm total length</td> </tr> <tr> <td>Kind of food</td> <td>Dry salmon diet (ASD2-30)</td> <td>Locally caught (live cut) earthworms</td> <td>Dry salmon diet (ASD2-30), live earth worms and brine shrimp nauplii</td> </tr> <tr> <td>Amount of food</td> <td>1g per tank in total.</td> <td>12-14 cm long earthworms were cut in half and distributed uniformly around the tank.</td> <td>Would not feed.</td> </tr> <tr> <td>Feeding frequency</td> <td>Not reported.</td> <td>Not reported.</td> <td>Would not feed.</td> </tr> <tr> <td>Pretreatment</td> <td>7-10 days acclimation period.</td> <td>7-10 days acclimation period.</td> <td>7-10 days acclimation period.</td> </tr> <tr> <td>Feeding of animals during test</td> <td>Fed 1 pellet of ASD2-30 at a time. Up to a total of 1 g per tank.</td> <td>Not reported.</td> <td>Would not feed.</td> </tr> </tbody> </table>			Criteria	Details			Species / strain	Brook trout <i>Salvelinus fontinalis</i>	Slimy Sculpin <i>Cottus cognatus</i>	Blacknose dace <i>Rhinichthys atratulus</i>	Source	Private hatchery	Local second-order streams	Local second-order streams	Wild caught	No	Yes – no other details given	Yes – no other details given	Age / size	Age not given 170 ± 11 mm total length	Age not given 70 ± 4 mm total length	Age not given 69 ± 4 mm total length	Kind of food	Dry salmon diet (ASD2-30)	Locally caught (live cut) earthworms	Dry salmon diet (ASD2-30), live earth worms and brine shrimp nauplii	Amount of food	1g per tank in total.	12-14 cm long earthworms were cut in half and distributed uniformly around the tank.	Would not feed.	Feeding frequency	Not reported.	Not reported.	Would not feed.	Pretreatment	7-10 days acclimation period.	7-10 days acclimation period.	7-10 days acclimation period.	Feeding of animals during test	Fed 1 pellet of ASD2-30 at a time. Up to a total of 1 g per tank.	Not reported.	Would not feed.
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Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (1 of 2)

3.4.3	Test system	Criteria	Details
		Test type	Flow through.
		Renewal of test solution	Flow through maintained at 6 L / min.
		Volume of test vessels	Volume of test vessels not reported, but water volume maintained at 85 L.
		Volume/animal	Not reported.
		Number of animals/vessel	20 Blacknose dace / vessel 10 Slimy sculpin / vessel 10 Brook trout / vessel
		Number of vessels/concentration	4 treatment levels of carbon dioxide with 3 replicates of each, giving 12 in total. Control tank: $0.0 \pm 0.0\%$ dissolved CO_2 Low: $1.4 \pm 0.4\%$ dissolved CO_2 Medium: $2.8 \pm 0.6\%$ dissolved CO_2 High: $5.1 \pm 1.3\%$ dissolved CO_2
		Test performed in closed vessels due to significant volatility of TS	Not reported.
3.4.4	Test conditions	Criteria	Details
		Test temperature	9°C – measurements conducted during test not actually reported, but largely unaffected.
		Dissolved oxygen	11.2 mg/L - measurements conducted during test not actually reported, but largely unaffected.
		pH	6.8 – during treatment, pH decreased to 5.5 except in the control vessels.
		Adjustment of pH	Not reported.
		Aeration of dilution water	Not reported.
		Intensity of irradiation	Not reported.
		Photoperiod	Not reported.
3.4.5	Duration of test	24 hours.	
3.4.6	Test parameter	Blood physiological variables (including hematocrit levels), behaviour including feeding responses before, during and after exposure.	
3.4.7	Sampling	Dissolved CO_2 concentrations measured throughout the 24 hour treatment period using a model GD-444 CEA instrument gas monitor with gas sampling pump.	
3.4.8	Monitoring of TS concentration	CO_2 levels monitored periodically during the 24-hour treatment period.	

**Section A7.4.1.1
Annex Point IIA7.1****Acute toxicity to fish (1 of 2)****4 RESULTS****4.1 Limit test**

Not performed.

4.1.1 Concentration

Not required.

4.1.2 Number / percentage of animals showing adverse effects

Not required.

4.1.3 Nature of adverse effect

Not required.

4.2 Results test substance

4.2.1 Initial concentration of substance

Not reported.

4.2.2 Actual concentration of test substance

The experiment was designed to cover the exposure to carbon dioxide at three levels: $1.4 \pm 0.4\%$ (low), $2.8 \pm 0.6\%$ (moderate) and $5.1 \pm 1.3\%$ (high).

4.2.3 Effect data (mortality)

Mortality was not being investigated in this study.

4.2.4 Concentration / response curve

Not relevant.

4.2.5 Other effects

Effects were observed on hematocrit levels, plasma glucose levels, ventilation rates, pectoral fin beats and cough rates.

4.3 Results of controls

4.3.1 Number / percentage of animals showing adverse effects

The number / percentage of animals showing adverse effects was not directly reported. However, the proportional (and percentage) of behavioural variables that showed post-treatment recovery to pre-treatment rates after exposure to various treatment levels of dissolved CO₂ is shown below:

Treatment level	Brook trout	Slimy sculpin	Blacknose dace	All species
Control		0/1 (0%)	1/1 (100%)	1/2 (50%)
Low	0/1 (0%)			0/1 (0%)
Medium	1/2 (50%)	1/2 (50%)	5/6 (83%)	7/10 (70%)
High	3/7 (43%)	4/5 (8%)	5/5 (100%)	12/17 (71%)
All levels	4/10 (40%)	5/8 (50%)	11/12 (92%)	20/30 (67%)

4.3.2 Nature of adverse effects

This study was not specifically looking for adverse effects. It was looking at behavioural effects such as ventilation rates, pectoral fin beats and cough rates. Other effects on hematocrit levels and plasma glucose were seen.

4.4 Test with reference substance

4.4.1	Concentration	Test not performed.
4.4.2	Results	Test not performed.

Section A7.4.1.1 **Acute toxicity to fish (1 of 2)**
Annex Point IIA7.1

5 APPLICANT'S SUMMARY AND CONCLUSION	
5.1	<p>Materials and methods</p> <p>This study was not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075.</p> <p>Rather than looking at acute toxicity <i>per se</i>, this investigates the physiological and behavioural effects of fish exposed to carbon dioxide.</p> <p>Brook trout, blacknose dace and slimy sculpin were tested in a system designed to stimulate an acute exposure to CO₂. Three replicates of four different CO₂ levels were tested in treatment vessels. Substrate cover of flat creekbed stones (5-15 cm) was provided in each tank; flow maintained at 6 L/minute and water volume at 85 L. Blacknose dace and slimy sculpin were collected from local second-order streams and brook trout from a private hatchery. After 2 days in laboratory holding tank, fish were anaesthetised, measured, sorted and distributed into tanks to give 20 blacknose dace, 10 slimy sculpin and 10 brook trout per tank. Numbers and weight of fish were designed to provide adequate samples of blood and behaviour without overcrowding.</p> <p>General water quality from source to experimental tanks was similar and all fish were allowed 7 – 10 days acclimation to test tanks during which they were observed for signs of parasites and other pathology. All fish were offered suitable diets, although blacknose dace would not feed throughout.</p> <p>Dissolved CO₂ was administered to 4 reservoir tanks from gas cylinders. In each reservoir tanks, 9°C well water was mixed with the CO₂ treated water to obtain test levels. The reservoir tanks then supplied each replicate tank.</p> <p>Percentage CO₂ readings, pH and other water quality variables were measured.</p> <p>All 3 fish species were exposed to treatment levels of control (0.0 ± 0.0%), low (1.4 ± 0.4%), medium (2.8 ± 0.6%), and high (5.1 ± 1.3%) dissolved CO₂ concentrations for 24 hours. CO₂ concentration was measured throughout the 24-hour period, and adjustments made periodically to maintain treatments at or near prescribed points. After tests, fish were monitored for a 1-week period to assess short-term mortality.</p>
5.2	<p>Results and discussion</p> <p>Physiological responses differed by species. All species had elevated hematocrits after 1 hour of exposure. Brook trout plasma glucose levels were raised after 1 hour. All species showed increased branchial ventilation, indicating stress although acclimation was indicated in blacknose dace after 24 hours. Brook trout had the longest reaction to stress at lower carbon dioxide levels.</p> <p>Recovery to pre-treatment activity rates of most behaviour patterns</p>

(including feeding) was observed 24 hours after cessation of exposure in all three species.

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.1	Acute toxicity to fish (1 of 2)	
Annex Point IIA7.1		

5.2.1	LC ₀	Not reported.
5.2.2	LC ₅₀	Not reported.
5.2.3	LC ₁₀₀	Not reported.
5.3	Conclusion	Rather than looking at acute toxicity <i>per se</i> , this test investigated the physiological and behavioural effects of 3 species of fish exposed to carbon dioxide. The results show that physiological responses to increased carbon dioxide in fish differed by species when they were exposed to 1.4%, 2.8% and 5.1% carbon dioxide. However, recovery to pre-treatment activity rates of most behaviour patterns (including feeding) was observed 24h after cessation of exposure in all 3 test species.
5.3.1	Other conclusions	None made.
5.3.2	Reliability	3
5.3.2	Deficiencies	Yes. This study was not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075. Rather than looking at acute toxicity <i>per se</i> , this investigates the physiological and behavioural effects of fish exposed to carbon dioxide. It should be noted that the use of carbon dioxide by Rentokil Initial would not increase the normal atmospheric concentrations of carbon dioxide in the locality.

Evaluation by Competent Authorities	
<i>Use separate "evaluation boxes" to provide transparency as to the comments and views submitted</i>	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Materials and methods	<i>Adopt applicant's version or include revised version. If necessary, discuss relevant discrepancies referring to the (sub) heading numbers and to applicant's summary and conclusion.</i>
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Reliability	<i>Based on the assessment of materials and methods include appropriate reliability indicator (the text in section 4.4.2.5.1 gives guidance on this point)</i>
Acceptability	<i>Acceptable / not acceptable</i> <i>(give reasons if necessary, e.g. if a study is considered acceptable despite a poor reliable indicator. Discuss the relevance of deficiencies and indicate if repeat is necessary.)</i>
Remarks	
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Reliability	<i>Discuss if deviating from view of rapporteur member state</i>
Acceptability	<i>Discuss if deviating from view of rapporteur member state</i>

Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (2 of 2)

		1 REFERENCE	Official use only
1.1	Reference	[REDACTED]	
1.2	Data protection	[REDACTED]	
1.2.1	Data owner	[REDACTED]	
1.2.3	Criteria for data protection	[REDACTED]	
		2 GUIDELINES AND QUALITY ASSURANCE	
2.1	Guideline study	No Not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075.	
2.2	GLP	No GLP was not compulsory at the time the study was performed.	
2.3	Deviations	Yes. This study did not follow the test guidelines of Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075. Rather than looking at acute toxicity <i>per se</i> , this report investigates the survival of the fish species at various concentrations of carbon dioxide.	
		3 MATERIALS AND METHODS	
3.1	Test material	As given in section 2.	
3.1.1	Lot/Batch number	Not reported.	
3.1.2	Specification	[REDACTED]	
3.1.3	Purity	[REDACTED]	

Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (2 of 2)

3.14	Composition of product	Not applicable for active substance.																															
3.15	Further relevant properties	Not reported.																															
3.16	Method of analysis	Not reported.																															
3.2	Preparation of TS solution for poorly soluble or volatile test substances	Not applicable. Carbon dioxide obtained and used from commercially available cylinder.																															
3.3	Reference substance	No.																															
3.3.1	Method of analysis for reference substance	Not applicable as reference substance was not used.																															
3.4	Testing procedure	3.4.1																															
3.4.1	Dilution water	<table border="1"> <thead> <tr> <th>Criteria</th> <th colspan="2">Details</th> </tr> </thead> <tbody> <tr> <td>Source</td> <td colspan="2">Tap water.</td> </tr> <tr> <td>Alkalinity</td> <td colspan="2">Not reported.</td> </tr> <tr> <td>Hardness</td> <td colspan="2">Not reported.</td> </tr> <tr> <td>pH</td> <td colspan="2">Not reported.</td> </tr> <tr> <td>Oxygen content</td> <td colspan="2">Experiments with trout designed to cover range of concentrations of dissolved oxygen from 0.5 to 10 ppm.</td> </tr> <tr> <td>Conductance</td> <td colspan="2">Not reported.</td> </tr> <tr> <td>Holding water different from dilution water</td> <td colspan="2">Not reported.</td> </tr> </tbody> </table>		Criteria	Details		Source	Tap water.		Alkalinity	Not reported.		Hardness	Not reported.		pH	Not reported.		Oxygen content	Experiments with trout designed to cover range of concentrations of dissolved oxygen from 0.5 to 10 ppm.		Conductance	Not reported.		Holding water different from dilution water	Not reported.							
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Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (2 of 2)

3.4.3	Test system	Criteria	Details
		Test type	Not reported.
		Renewal of test solution	Not reported.
		Volume of test vessels	40 litres.
		Volume/animal	Not reported.
		Number of animals/vessel	10 trout or 5 trout and 5 perch per vessel.
		Number of vessels/concentration	3 vessels at 12.5°C, 16.5°C and 19.5°C. Same quantity of gas in each: 0.5 – 10 ppm dissolved oxygen and 0 – 240 ppm carbon dioxide.
		Test performed in closed vessels due to significant volatility of TS	Yes. Vessels sealed from atmosphere by a glass plate positioned on top.
3.4.4	Test conditions	Criteria	Details
		Test temperature	12.5°C, 16.5°C and 19.5°C ± 0.5°C.
		Dissolved oxygen	0.5 – 10 ppm ± 5°C.
		pH	Not reported.
		Adjustment of pH	Not reported.
		Aeration of dilution water	Not reported.
		Intensity of irradiation	Not reported.
		Photoperiod	Not reported.
3.4.5	Duration of test	Up to 24 hours (if any fish remained alive).	
3.4.6	Test parameter	Mortality.	
3.4.7	Sampling	Not reported.	
3.4.8	Monitoring of TS concentration	Carbon dioxide concentration estimated by determining the pH value, temperature and bicarbonate alkalinity of a sample and calculating the free carbon dioxide concentration using predetermined nomograms.	
3.4.9	Statistics	Standard deviations and means of the logarithms of periods of survival were assessed.	
4 RESULTS			
4.1	Limit test		
4.1.1	Concentration	Test not performed.	
4.1.2	Number / percentage of animals showing adverse effects	Test not performed.	
4.1.3	Nature of adverse effect	Test not performed.	

Section A7.4.1.1
Annex Point IIA7.1
Acute toxicity to fish (2 of 2)

4.2	Results test substance	
4.2.1	Initial concentration of substance	Not reported, however the experiment was designed to cover a range of concentrations of carbon dioxide from 0 – 240 ppm.
4.2.2	Actual concentration of test substance	Not reported, however the experiment was designed to cover a range of concentrations of carbon dioxide from 0 – 240 ppm.
4.2.3	Effect data (mortality)	All fish died within 24 hours, ie 100% mortality.
4.2.4	Concentration / response curve	Not reported.
4.2.5	Other effects	Not reported.
4.3	Results of controls	
4.3.1	Number / percentage of animals showing adverse effects	Not reported.
4.3.2	Nature of adverse effects	Not reported.
4.4	Test with reference substance	
4.4.1	Concentration	Test not performed.
4.4.2	Results	Test not performed.
5 APPLICANT'S SUMMARY AND CONCLUSION		
5.1	Materials and methods	<p>This study was not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075. Rather than looking at acute toxicity <i>per se</i>, this report investigates the survival of the fish species at various concentrations of carbon dioxide.</p> <p>The experiments were designed to cover a range of concentrations of 0 - 240 ppm carbon dioxide at temperatures of 12.5°C, 16.5°C and 19.5°C.</p> <p>The desired concentrations of oxygen and carbon dioxide were obtained by passing through the test aquarium mixtures of nitrogen, oxygen and carbon dioxide in suitable proportions at a rate of about 50ml/second (monitored by a flow meter). This rate ensured that there were no appreciable alterations in the concentrations of oxygen and carbon dioxide due to the respiration of the fish. The three aquarium were filled with tap water and the tops sealed from the atmosphere with glass plates so that the surface of the water was in contact with the gas mixture (the excess of which was led to waste through a small exhaust vent). The temperature of the aquariums was maintained within $\pm 0.5^\circ\text{C}$.</p> <p>Concentrations of dissolved oxygen and carbon dioxide were monitored throughout the tests. Fish in groups of 10 were tested in each aquarium (10 trout or 5 trout and 5 perch). They were sorted into batches at random</p>

and acclimatised at the appropriate temperature without food for 24 hours prior to testing. The period of survival for each fish was recorded as minutes between immersion of the fish in a test aquarium and the time at which all movements, respiratory and otherwise, had ceased.

Rentokil Initial plc

Carbon Dioxide

March 2004

Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (2 of 2)

5.2	Results and discussion	It is shown that concentrations of carbon dioxide that sometimes occur in polluted streams can more than double the minimum concentration of dissolved oxygen necessary for the survival of half a population of rainbow trout fingerlings for 24 hours. Increase in temperature between 12.5°C and 19.5°C shortens period of survival in solutions containing up to 67 ppm carbon dioxide.
5.2.1	LC ₀	Not reported.
5.2.2	LC ₅₀	Not reported.
5.2.3	LC ₁₀₀	Not reported.
5.3	Conclusion	Not reported.
5.3.1	Other conclusions	Not reported
5.3.2	Reliability	3
5.3.2	Deficiencies	Yes This study was not carried out to Guideline C.1 in Annex V of Directive 67/548/EEC, to OECD Guideline 203, or to US-EPA guideline OPPTS 850.1075. Rather than looking at acute toxicity <i>per se</i> , this report investigates the survival of the fish species at various concentrations of carbon dioxide. It should be noted that the use of carbon dioxide by Rentokil Initial would not increase the normal atmospheric concentrations of carbon dioxide in the locality

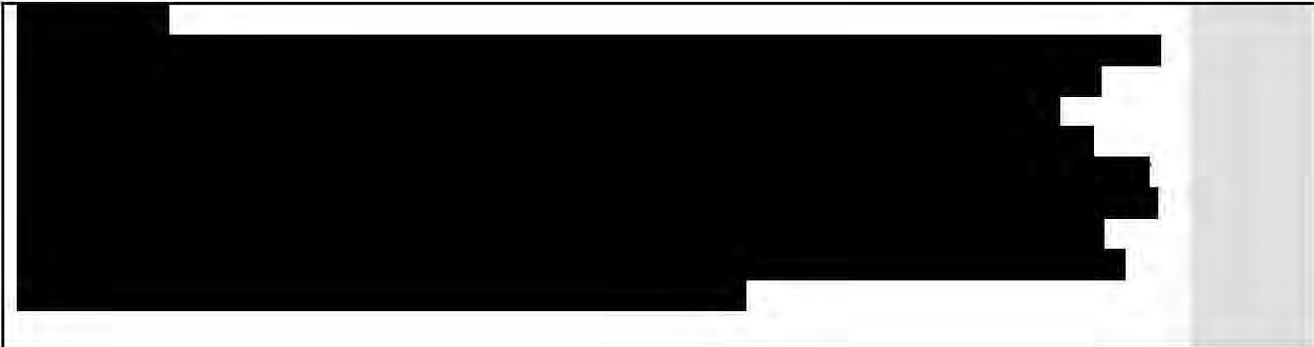
Section A7.4.1.1
Annex Point IIA7.1

Acute toxicity to fish (2 of 2)

Evaluation by Competent Authorities	
	<i>Use separate "evaluation boxes" to provide transparency as to the comments and views submitted</i>
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Materials and methods	<i>Adopt applicant's version or include revised version. If necessary, discuss relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion.</i>
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COMMENTS FROM	
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Results and discussion	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Reliability	<i>Discuss if deviating from view of rapporteur member state</i>
Acceptability	<i>Discuss if deviating from view of rapporteur member state</i>

Table 4-2: Standard form for justification of the non-submission of data

<p>Section A7.4.1.2 Annex Point/TNsG Annex IIA, VII 7.2</p>	<p>Acute toxicity to invertebrates Section 7: Ecotoxicological Profile, including Fate and Behaviour</p>	
<p>JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		<p>Official use only</p>
<p>Other existing data</p>	<p><input checked="" type="checkbox"/> Technically not feasible []</p>	<p>Scientifically unjustified []</p>
<p>Limited exposure</p>	<p><input checked="" type="checkbox"/> Other justification []</p>	
<p>Detailed justification:</p>	<p>An acute toxicity study for carbon dioxide to invertebrates is not considered necessary for the following reasons:</p> <ol style="list-style-type: none"> 1. It is not scientifically necessary because under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water, outside normal atmospheric ranges. <i>See next page for full details of the scientific calculation, which supports this statement.</i> 2. There are two studies available in the public domain which investigates the effects of carbon dioxide to <i>Daphnia magna</i> and aquatic cladocera species, including <i>Daphnia similis</i>. This study shows that <i>Daphnia similis</i> could tolerate carbon dioxide levels of 86 mg/L in water (86 ppm, or 0.0086%). <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i> could tolerate 12.0 mg/L (12 ppm or 0.0012 %) and 32.6 mg/L (32.6 ppm or 0.0326 mg/L). <i>Daphnia magna</i> can tolerate levels of carbon dioxide up to 2% v/v. The carbon dioxide values cited here are naturally occurring exposures. <i>Refer to attached study summary for details about the data available on the toxicity of carbon dioxide to aquatic invertebrates.</i> <p>Continued....</p>	



Section A7.4.1.2 Annex Point/TNsG Annex IIA, VII 7.2	Acute toxicity to invertebrates Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:
(continued):

[REDACTED]

[REDACTED]

Given that substantial rises in atmospheric carbon dioxide is never going to be reached under the normal conditions of use of Rentokil Initial's biocidal products and the data cited in this application, it is scientifically unjustified to conduct further studies.

[REDACTED]

[REDACTED]

[REDACTED]

Undertaking of intended	Not applicable
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<p>Section A7.4.1.2 Annex Point/TNsG Annex IIA, VII 7.2</p>	<p>Acute toxicity to invertebrates Section 7: Ecotoxicological Profile, including Fate and Behaviour</p>
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<p>Evaluation by Competent Authorities</p>	
	<p>Use separate “evaluation boxes” to provide transparency as to the Comments and views submitted</p>
	<p>EVALUATION BY RAPPORTEUR MEMBER STATE</p>
<p>Date</p>	<p><i>Give date of action</i></p>
<p>Evaluation of applicant’s justification</p>	<p><i>Discuss applicant’s justification and, if applicable, deviating view</i></p>
<p>Conclusion</p>	<p><i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i></p>
<p>Remarks</p>	
	<p>COMMENTS FROM OTHER MEMBER STATES (specify)</p>
<p>Date</p>	<p><i>Give date of comments submitted</i></p>
<p>Evaluation of applicant’s justification</p>	<p><i>Discuss if deviating from view of rapporteur member state</i></p>
<p>Conclusion</p>	<p><i>Discuss if deviating from view of rapporteur member state</i></p>
<p>Remarks</p>	

Section A7.4.1.2

Acute toxicity to invertebrates (1 of 3)

Annex Point IIA, VII, 7.2

Special investigation in *Daphnia magna*Official
use only**1. REFERENCE****1.1 Reference**

[REDACTED]

1.2 Data protection

[REDACTED]

1.2.1 Data owner

[REDACTED]

1.2.2

1.2.3 Criteria for data protection

[REDACTED]

2. GUIDELINES AND QUALITY ASSURANCE**2.1 Guideline study**

No.
Not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.

2.2 GLP

No.
No information available on whether study complies with the requirements of GLP.

2.3 Deviations

Yes.
No set guideline followed.

3. MATERIALS AND METHODS**3.1 Test material**

As given in section 2.

3.1.1. Lot/Batch number

Not reported.

3.1.2 Specification

[REDACTED]

[REDACTED]

3.2 Preparation of TS solution for poorly soluble or volatile test substances

Not applicable. No special procedures were applied to determine carbon dioxide content in water

3.3 Reference substance

No.

3.3.1 Method of analysis for reference substance

Reference substance not used.

3.4 Testing procedure

3.4.1 Dilution water

1:14 diluted seawater.
No other information reported.

3.4.2 Test organisms

Female *Daphnia magna* Straus
Refer to table A7_1_1_2-1 for description of test organisms.

3.4.3 Test system

Refer to table A7_1_1_2-2 for description of test system.

Section A7.4.1.2	Acute toxicity to invertebrates (1 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in <i>Daphnia magna</i>	

3.4.4	Test conditions	Refer to table A7_1_1_2-3 for description of test conditions.
3.4.5	Duration of test	30 minutes.
3.4.6	Test parameter	Heart rate, and pH changes.
3.4.7	Sampling	Refer to table A7_1_1_2-3 for description of test conditions.
3.4.8	Monitoring of TS concentration	Refer to table A7_1_1_2-3 for description of test conditions.
3.4.9	Statistics	Not reported.
4 RESULTS		
4.1	Limit Test	Not performed.
4.2	Results test substance	
4.2.1	Initial concentrations of test substance	Carbon dioxide concentrations were increased to 2% (oxygen concentration were either at normal levels, or decreased to 3.6%). Note that author comments that <i>Daphnia</i> are known to be naturally exposed to ambient carbon dioxide concentrations of up to 2%.
4.2.2	Actual concentrations of test substance	Carbon dioxide concentrations were increased to 2% (oxygen concentration were either at normal levels, or decreased to 3.6%). Note that author comments that <i>Daphnia</i> are known to be naturally exposed to ambient carbon dioxide concentrations of up to 2%.
4.2.3	Effect data (Immobilisation)	Heart rate changes could not be provoked by P _{CO2} changes during normoxia (normal levels of oxygen). However, during severe hypoxia (low oxygen concentration of 3.6%), the application of hypercapnia (2% carbon dioxide, pH 6) caused a decrease of heart rate of <i>Daphnia magna</i> by 20-50 beats per minute. The minimum was reached within a few minutes. After 20 minutes, the heart had returned back to a rate similar to the pre-hypercapnic value (the normocapnic value at pH 7.45). Switching then to normocapnia or acapnia (pCO ₂ : 0%, pH 8.5) caused the heart rate either to non-transiently or transiently to increase. Concerning blood pH, the application of hypercapnia (2% carbon dioxide, pH 6) caused a transient decrease of pH even during normoxia. The minimum was reached within 10 minutes. After 20-30 minutes the pH had increased to a stable value which was a little lower than the pre-hypercapnic one.
4.2.4	Concentration/response curve	Not reported.
4.2.5	Other effects	None reported.
4.3	Results of controls	No control species reported.
4.4	Test with reference substance	Not performed.
5.1	Materials and Methods	5 APPLICANT'S SUMMARY AND CONCLUSION This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC. (Continued...)

Section A7.4.1.2

Acute toxicity to invertebrates (1 of 3)

Annex Point IIA, VII, 7.2

Special investigation in *Daphnia magna*

5.1 Materials and Methods

(Continued)

Female individuals of *Daphnia magna* Straus were used. In most experiments, the animals had a body length of 2.5 +/- 0.2 mm (measured from the anterior most part of the head to the base of the apical spine), and a body mass of approximately 1 mg. The animals descended from a clone (clone 5) cultured at the University of Sheffield, Department of Animal and Plant Physiology, United Kingdom.

An animal chamber made of anodized aluminium was utilised for the experiments. Two cover slides were used as top and bottom of the inner room, whose lateral wall was the inside of the water filled thermostated casing. A constant medium flow (8-10 ml min⁻¹) through the chamber was generated by a peristaltic pump located after it. The test solution (1:14 diluted seawater) was sucked up from a thermostated glass vessel with a small opening (transport time from vessel to chamber 6s), where it was equilibrated with different O₂/N₂ mixtures (at normocapnic conditions 400 ppm CO₂), using gas mixing pumps. Temperature and oxygen content of the medium leaving the chamber was continually checked using a needle-shaped thermoelement (NiCr-Ni Sensortek, Clifton NJ) and a small oxygen electrode (P/N SI 130, Strathkelvin Instruments Glasgow, UK).

Single *Daphnia magna* were fixed in the chamber by gently screwing the chamber's top (which has a fine thread) down. The animals were acclimated in the chamber for about 1h. Within the first 30 minutes of acclimation the heart rate usually reached a constant level. At the beginning of the actual experiment the animals were further exposed to normoxia (normal levels of oxygen) / normocapnia (normal levels of carbon dioxide) for 15-20 minutes. Then, after switching the gas-mixing pump the medium in the thermostated glass vessel was equilibrated with nitrogen (at normal levels of carbon dioxide) to remove oxygen (Anoxia: oxygen levels < 0.06 %). Anoxia was not instantaneously reached in the medium perfusing the animal chamber, and a short hypoxic period (where there was a deficiency in the amount of oxygen reaching body tissues) occurred for approximately 10 minutes. After the experiments on the physiological responses to short or long-term anoxia (absence of oxygen), normal levels of oxygen (normoxia) was applied again.

Heart rate was evaluated by video microscopy and digital image processing, utilising the rhythmic variations in pixel intensity. The amplitude of heart wall movements in fixed animals, being a relative measure of stroke volume, was evaluated utilising video recordings. During the experiments on circulation, the medium's temperature was 15°C.

The transparency of *Daphnia magna* allowed the application of pH sensitive dyes to determine pH changes. The diluted dyes were injected into the blood capillaries (1B100F-4, WPI Sarasota FL) were pulled (micropipette puller from Zeschka Zoological Institute of the University of Munich Germany) and bevelled (beveler 1300M, WPI Sarasota) to an angle of 30° between capillary and a

(Continued...)

Section A7.4.1.2

Acute toxicity to invertebrates (1 of 3)

Annex Point IIA, VII, 7.2

Special investigation in *Daphnia magna*

5.2 Materials and Methods

(Continued)

0.3 µm aluminium oxide coated film (3 M Neuss, Germany), resulting in a diameter of the opening of 9–10 µm. The pointed shape made penetration of the cuticle easier. Micropipette movements were controlled via a motor driven micromanipulator (AM3 DC-K, control unit BA-ST 3, Bachofer Reutlingen Germany). Injection pressure was provided by a microinjector (Transjector 5246, Eppendorf Hamburg Germany). During injection, the animals (being in a small droplet of medium) became shortly immobilised, because the micropipette pressed them lightly to a semi-circular silicone support on a microscope slide. Another technique was to briefly fix the apical spine to the slide using wax. For that, the medium around the spine was shortly removed. Approximately 10 ml of dye solution (which is approximately 1.5% of total blood space) were injected into a dorsal lacuna carrying blood directly to the heart. Direct observation of the heart showed that such small injected volumes did not cause any significant heart rate changes. After the experiments, the animals showed normal swimming patterns. In control experiments, the animals tolerated injection volumes ten times higher.

Blood pH changes were measured at temperatures of 20°C using the pH sensitive fluorescent dye BCECF (Molecular Probes Eugene OR). A stock solution of BCECF in dimethyl sulphoxide was stored at -20°C. Before injecting, the stock solution was diluted with Ringer's solution to a final concentration of 1 mM BCECF. Monochromatic light was generated using a rapid scanning monochromator equipped with xenon arc lamp (TILL Phototonics, Planegg Germany). The fluorescence image (of the anterior body part of *Daphnia*) was detected with a liquid-nitrogen cooled CCD camera which took 2 frames per second (LN/CCD-576E Princeton Instruments Trenton NJ). As dye concentration in different body parts may vary, the measurements were done in excitation ratio mode (excitation 439 nm and 490 nm, emission measured above 520 nm). Being interested in pH changes and not absolute values, time-consuming calibration procedures were not applied.

Confocal laser microscopy was used to image the extra and intracellular distribution of pH sensitive dyes in *Daphnia*, and pH changes during anoxia (oxygen deficiency) was studied (at 20°C). Two different kinds of the pH sensitive fluorescent dye SNARF-1 (molecular probes) were used for the emission ratio measurements (excitation at 488 nm, emissions measured at 588 and 604 nm) (i) to image blood spaces only and to prevent the penetration of dye into cells, the dextran-coupled indicator (dextran, SNARF-1, 70,000 MW anionic), was applied. (ii) for an imaging of intracellular spaces the cell-permanent ester (SNARF calcein AM) was injected. In both cases approximately 10 ml of dye solution (dye dissolved in 1:3 diluted sea water) was used. The experimental conditions were in principle the same as described above, with the exception that normocapnia (normal levels of carbon dioxide) could not be maintained during anoxia (absence of oxygen), as gas-mixing pumps were not available at that time. But these experiments and the BCECF measurements at anoxia/normocapnia yielded similar results. They have been included because they demonstrate clearly the advantages of optical methods to image systemic processes.

5.2 Results and discussion

Heart rate and pH changes at varying ambient P_{CO_2} / pH were studied in *Daphnia magna*. Gas mixing pumps were utilised to modify P_{CO_2} in the medium, resulting also in pH changes. Heart rate changes could not be provoked by P_{CO_2} changes during normoxia (normal levels of oxygen). However, during severe hypoxia (approximately 3.6% oxygen), the application of hypercapnia (2% carbon dioxide, pH 6) caused a transient decrease of heart rate by 20-50 beats per minute. The minimum was reached within a few minutes. After 20 minutes, the heart had returned back to a rate similar to the pre-hypercapnic (normocapnic) value (pH 7.45). Switching then to normocapnia (normal levels of carbon dioxide) or acapnia (0% carbon dioxide, pH 8.5) caused the heart rate to either non-transiently or transiently to increase. Concerning blood pH, the application of hypercapnia (increased carbon dioxide) caused a transient decrease of pH even during normoxia (normal levels of oxygen). The minimum was reached within 10 minutes. After 20-30 minutes, the pH had increased to a stable value, which was a little lower than the pre-hypercapnic one.

The pH measurements by optical methods in whole animals are novel, and should not yet be regarded as a final analysis of acid-base balance in *Daphnia*, but they give a first view on specific links between pH, heart activity and metabolism. The intracellular pH measured in the shell gland reflected the anaerobic metabolic activity. Lactate production is indirectly linked with proton release, and anoxia (absence of oxygen) caused intracellular pH to decrease. Due to protons released from the cells, the extracellular pH also dropped due to anoxia showing a time-course which is a little faster, but similar to the heart rate course. During early anoxia lactate and protons were generated at a high rate in cells and tissues, and they were released to the blood and also transported to the heart. The similarity of both time-courses, heart rate and pH indicates that heart activity (heart rate) is influenced or even controlled by pH. Steady changes in carbon dioxide partial pressure / pH caused a transient change in blood pH. *Daphnia magna* was obviously able to control pH during an ambient-causes respiratory acidosis. A steady change of ambient carbon dioxide partial pressure / pH caused a transient drop of heart rate (with a time course similar to blood pH) indicating again a functional relationship between heart rate and pH. In *Daphnia* the control of respiration depends on adaptive changes of heart rate. Apart from ambient oxygen partial pressure, internal carbon dioxide pressure and pH may also be input signals for respiratory control. It is known, in any case, that a decreasing extracellular or intracellular pH causes a depression of trans-sarcolemmal Ca^{2+} currents and contraction in cardiac muscle cells.

5.2.1 EC_0 Not reported.

5.2.2 EC_{50} Not reported.

5.2.3 EC_{100} Not reported.

5.3 Conclusion

Data reported in this study shows that *Daphnia magna* can tolerate carbon dioxide partial pressures of 2 % v/v both in the presence of normal levels of oxygen and severe hypoxia (3.6% oxygen).

Note that author comments that *Daphnia* are known to be naturally

exposed to ambient carbon dioxide concentrations of up to 2%.

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (1 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in <i>Daphnia magna</i>	

5.3.1	Reliability	3
5.3.2	Deficiencies	<p>Yes.</p> <p>This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.</p> <p>Rather than looking at acute toxicity <i>per se</i>, this report investigates heart rate and pH changes at varying ambient P_{CO_2} / pH in <i>Daphnia magna</i>. This study shows that <i>Daphnia magna</i> can tolerate carbon dioxide partial pressures of 2 % v/v both in the presence of normal levels of oxygen and severe hypoxia (3.6% oxygen).</p> <p>It should be noted that the use of carbon dioxide by Rentokil Initial would not increase the normal atmospheric concentrations of carbon dioxide in the locality.</p>

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2 Annex Point IIA, VII, 7.2	Acute toxicity to invertebrates (1 of 3) Special investigation in <i>Daphnia magna</i>	

Table A7_1_1_2-1 Description of test organisms

Criteria	Details
Strain	<i>Daphnia magna</i> Straus
Source	Descended from a clone (clone 5) cultured at the University of Sheffield, Department of Animal and Plant Physiology, United Kingdom.
Age	Not reported, but in most experiments the animals had a body length of 2.5 +/- 0.2 mm (measured from the anterior most part of the head to the base of the apical spine), and a body mass of approximately 1 mg.
Breeding method	Descended from a clone (clone 5) cultured at the University of Sheffield, Department of Animal and Plant Physiology, United Kingdom.
Kind of food	Algae
Feeding frequency	Not fully described, other than "keeping and feeding of animals were identical to previous studies by the same author."
Pre-treatment	Animals were acclimated in the chamber for about 1h.
Feeding of animals during test	Not fully described, other than "keeping and feeding of animals were identical to previous studies by the same author."

Table A7_1_1_2-2 Description of test system

Criteria	Details
Renewal of test solution	A constant medium flow (8-10 ml min ⁻¹) through the chamber was generated by a peristaltic pump located after it. The test solution (1:14 diluted seawater) was sucked up from a thermostated glass vessel with a small opening (transport time from vessel to chamber 6s), where it was equilibrated with different O ₂ /N ₂ mixtures (at normocapnic conditions 400 ppm CO ₂), using gas mixing pumps.
Volume of test vessels	Not reported. Test vessels described as follows: An animal chamber made of anodized aluminium was utilised for the experiments. Two cover slides were used as top and bottom of the inner room, whose lateral wall was the inside of the water filled theromstated casing.
Volume/animal	Not reported.
Number of animals/vessel	One.
Number of vessels / concentration	Not reported.

Test performed in closed vessels due to significant volatility of TS	Yes.
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Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (1 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in <i>Daphnia magna</i>	

Table A7_1_1_2-3 Description of test conditions

Criteria	Details
Test temperature	Temperature of the medium leaving the chamber (see table A7_1_1_2-2 renewal of test solution) was continually checked using a needle-shaped thermoelement (NiCr-Ni Sensortek, Clifton NJ)
Dissolved oxygen	Oxygen partial pressure of the medium leaving the chamber (see table A7_1_1_2-2 renewal of test solution) was continually checked using a small oxygen electrode (P/N SI 130, Strathkelvin Instruments Glasgow, UK).
pH	The transparency of <i>Daphnia magna</i> allowed the application of pH sensitive dyes to determine pH changes.
Adjustment of pH	No.
Aeration of dilution water	Not reported.
Quality / intensity of irradiation	Not reported.
Photoperiod	Not reported.

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (1 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in <i>Daphnia magna</i>	

Evaluation by Competent Authorities	
	Use separate "evaluation boxes" to provide transparency as to the comments and views submitted.
	EVALUATION BY RAPPORTEUR MEMBER STATE
Date	<i>Give date of action</i>
Materials and Methods	<i>State if applicants version is acceptable, or indicate relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion.</i>
Results and discussion	<i>Adopt applicant's version or include revised version. If necessary, discuss relevant deviations from applicant's view referring to the (sub)heading numbers.</i>
Conclusion	Other conclusions: <i>(adopt applicant's version or include revised version)</i>
Reliability	<i>Based on assessment of materials and methods include appropriate reliability indicator.</i>
Acceptability	acceptable / not acceptable <i>(give reasons if necessary e.g. if a study is considered acceptable despite a poor reliability indicator. Discuss the relevance of deficiencies and indicate if repeat if necessary).</i>
Remarks	
	COMMENTS FROM
Date	<i>Give date of comments submitted.</i>
Materials and Methods	<i>Discuss additional relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion</i> <i>Discuss if deviating from view of rapporteur member state. .</i>
Results and discussion	<i>Discuss if deviating from view of rapporteur member state.</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state.</i>
Reliability	<i>Discuss if deviating from view of rapporteur member state.</i>
Acceptability	<i>Discuss if deviating from view of rapporteur member state.</i>
Remarks	

Section A7.4.1.2

Annex Point IIA, VII, 7.2

Acute toxicity to invertebrates (2 of 3)

Special investigation in Cladocera species *Daphnia similis*
Momia flagellata and *Ceriodaphnia rigaudi*

		1. REFERENCE	Official use only
1.1	Reference	[REDACTED]	
1.2	Data protection	[REDACTED]	
1.2.1	Data owner	[REDACTED]	
1.2.2			
1.2.3	Criteria for data protection	No data protection claimed.	
		2. GUIDELINES AND QUALITY ASSURANCE	
2.1	Guideline study	No. Not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.	
2.2	GLP	No. GLP was not compulsory at the time the study was performed.	
2.3	Deviations	Yes. No set guideline followed.	
		3. MATERIALS AND METHODS	
3.1	Test material	As given in section 2.	
3.1.1.	Lot/Batch number	[REDACTED]	
3.1.2	Specification	[REDACTED]	
		[REDACTED]	
3.2	Preparation of TS solution for poorly soluble or volatile test substances	Not applicable. No special procedures were applied to determine carbon dioxide content in water.	
3.3	Reference substance	No.	
3.3.1	Method of analysis for reference substance	Reference substance not used.	

Section A7.4.1.2**Annex Point IIA, VII, 7.2****Acute toxicity to invertebrates (2 of 3)**

Special investigation in Cladocera species *Daphnia similis*
Monia flagellata and *Ceriodaphnia rigaudi*

3.4	Testing procedure	
3.4.1	Dilution water	Refer to table A7_1_1_2-1 for description of test site and sampling protocol
3.4.2	Test organisms	Abundance of cladocera in the test pond was used as indication of the effect of seasonal variation in carbon dioxide concentration. Species counted were <i>Daphnia similis</i> <i>Ceriodaphnia rigaudi</i> <i>eriodaphnia rigaudi</i>
3.4.3	Test system	Refer to table A7_1_1_2-1 for description of test site and sampling protocol
3.4.4	Test conditions	Refer to table A7_1_1_2-1 for description of test site and sampling protocol
3.4.5	Duration of test	12 months (measurements taken from July 1980 to June 1981).
3.4.6	Test parameter	Number of individuals present in test pond.
3.4.7	Sampling	Fortnightly. Refer to table A7_1_1_2-1 for description of test site and sampling protocol.
3.4.8	Monitoring of TS concentration	Monthly.
3.4.9	Statistics	Not reported.
		4 RESULTS
4.1	Limit Test	Not performed.
4.2	Results test substance	
4.2.1	Initial concentrations of test substance	Carbon dioxide fluctuated from almost nil to 86 mg/L during the period of investigation.
4.2.2	Actual concentrations of test substance	Carbon dioxide fluctuated from almost nil to 86 mg/L during the period of investigation.
4.2.3	Effect data (Immobilisation)	Carbon dioxide did not seem to be the conditioning factor for the production and growth of Cladocera. <i>Daphnia similis</i> not only occurred at carbon dioxide concentrations of 86 mg/L but registered a peak showing that they can survive in high carbon dioxide concentrations. <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i> showed peaks at carbon dioxide concentrations of 12 and 32.6 mg/L respectively.
4.2.4	Concentration/response curve	Not reported.
4.2.5	Other effects	None reported.
4.3	Results of controls	No control species reported.
4.4	Test with reference substance	Not performed.
		5 APPLICANT'S SUMMARY AND CONCLUSION
5.1	Materials and Methods	This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC. (Continued...)

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2 Annex Point IIA, VII, 7.2	Acute toxicity to invertebrates (2 of 3) Special investigation in Cladocera species <i>Daphnia similis</i> <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i>	
5.1 Materials and Methods (Continued)	<p>Water samples were collected from a pond, about 24 km from Ludhiana at Village Raqba (Ludhiana) situated at the link road connecting Boparai to Mullanpur. (India).</p> <p>The water level in pond varied from 20 cm in June to 40-50 cm in July – September. The water was muddy during June-July, greenish during August-September and transparent during February-March. The pH ranged between 7.20 (July) and 9.75 (December). The sample collections were made at fortnightly intervals by sieving 100 litres of water through a plankton net (300 meshes per square cm), from three marked points (A, B and C) at the peripheral regions of the pond. The material caught was preserved in 10% formalin. In the laboratory, the numerical estimation of population size was carried out according to Welch (1952).</p> <p>The water temperature, Secchi Disc Transparency, Colour and water depth were all measured before collecting the material. Dissolved oxygen, pH and carbon dioxide were estimated using standard methods for the examination of water and waste water (14th ed. Amer. Public Health Assoc Inc, New York).</p>	
5.2 Results and discussion	<p>The test pond harboured insects, fish, frogs and tortoises, besides plankton. The zooplankton largely comprised of Cladocera, Copepoda and Rotifera. Cladocerans were represented by six species in all, of these <i>Monia flagellata</i> was noted in maximum number of 15 August 1980 when its actual number was counted 1827 individuals per litre. This species was not found in the test pond from 15 November 1980 to 15 May 1981 when only 2 individuals per litre occurred. <i>Ceriodaphnia rigaudi</i> was found in the pond from 15 August 1980 to 15 May 1981 reaching maximum number on 15 September 1980. <i>Daphnia similis</i> was the third species found in abundance and occurred in the pond from 1 November 1980 to 15 May 1981, but was absent on 15 December 1980. The maximum pulse of this species was recorded on 15 January 1981, when its number reached 421 individuals per litre.</p>	
5.2.1 EC ₀	Not reported.	
5.2.2 EC ₅₀	Not reported.	
5.2.3 EC ₁₀₀	Not reported.	
5.3 Conclusion	<p>Data reported in this study shows that <i>Daphnia similis</i> not only occurred at carbon dioxide concentrations of 86 mg/L but registered a peak showing that they can survive in high carbon dioxide concentrations. <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i> showed peaks at carbon dioxide concentrations of 12 and 32.6 mg/L respectively.</p>	
5.3.1 Reliability	3	
5.3.2 Deficiencies	<p>Yes.</p> <p>This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.</p> <p>Rather than looking at acute toxicity <i>per se</i>, this report measures the natural fluctuations in background carbon dioxide concentrations in a pond, over 12 months and its effect on the survival of the Cladocera species <i>Daphnia similis</i> <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i>.</p> <p>It should be noted that the use of carbon dioxide by Rentokil Initial would not increase the normal atmospheric concentrations of carbon dioxide in the locality.</p>	

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (2 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in Cladocera species <i>Daphnia similis</i> <i>Monia flagellata</i> and <i>Ceriodaphnia rigaudi</i>	

Table A7_1_1_2-1 Description of test site and sampling protocol

Criteria	
Description of site	Water samples were collected from a pond, about 24 km from Ludhiana at Village Raqba (Ludhiana) situated at the link road connecting Boparai to Mullanpur. (India)
Description of test pond : Appearance of water	Water level in pond varied from 20 cm in June to 40-50 cm in July – September. The water was muddy during June-July, greenish during August-September and transparent during February-March.
Description of test pond : pH of water	pH ranged between 7.20 (July) and 9.75 (December).
Number of sampling sites	Three marked points (A, B and C) at the peripheral regions of the pond.
Sample intervals	Fortnightly
Sample preparation	100 litres of water sieved through a plankton net (300 meshes per square cm), and material caught was preserved in 10% formalin.

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2 Annex Point IIA, VII, 7.2	Acute toxicity to invertebrates (2 of 3) Special investigation in Cladocera species <i>Daphnia similis</i> <i>Momia flagellata</i> and <i>Ceriodaphnia rigaudi</i>	








Evaluation by Competent Authorities	
	Use separate "evaluation boxes" to provide transparency as to the comments and views submitted.
	EVALUATION BY RAPPORTEUR MEMBER STATE
Date	<i>Give date of action</i>
Materials and Methods	<i>State if applicants version is acceptable, or indicate relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion.</i>
Results and discussion	<i>Adopt applicant's version or include revised version. If necessary, discuss relevant deviations from applicant's view referring to the (sub)heading numbers.</i>
Conclusion	Other conclusions: <i>(adopt applicant's version or include revised version)</i>
Reliability	<i>Based on assessment of materials and methods include appropriate reliability indicator.</i>
Acceptability	acceptable / not acceptable <i>(give reasons if necessary e.g. if a study is considered acceptable despite a poor reliability indicator. Discuss the relevance of deficiencies and indicate if repeat if necessary).</i>
Remarks	
	COMMENTS FROM
Date	<i>Give date of comments submitted.</i>
Materials and Methods	<i>Discuss additional relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion</i> <i>Discuss if deviating from view of rapporteur member state. .</i>
Results and discussion	<i>Discuss if deviating from view of rapporteur member state.</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state.</i>
Reliability	<i>Discuss if deviating from view of rapporteur member state.</i>
Acceptability	<i>Discuss if deviating from view of rapporteur member state.</i>
Remarks	

Section A7.4.1.2

Acute toxicity to invertebrates (3 of 3)

Annex Point IIA, VII, 7.2

Special investigation in ants

		1. REFERENCE		Official use only
1.1	Reference			
1.2	Data protection			
1.2.1	Data owner			
1.2.2				
1.2.3	Criteria for data protection			
		2. GUIDELINES AND QUALITY ASSURANCE		
2.1	Guideline study	No. Not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.		
2.2	GLP	No. No record of study being conducted in compliance with the requirements of GLP.		
2.3	Deviations	Yes. No set guideline followed.		
		3. MATERIALS AND METHODS		
3.1	Test material	As given in section 2.		
3.1.1.	Lot/Batch number			
3.1.2	Specification			
				
3.2	Preparation of TS solution for poorly soluble or volatile test substances	Not applicable. No special procedures were applied to determine carbon dioxide content in water.		
3.3	Reference substance	No.		

Section A7.4.1.2**Acute toxicity to invertebrates (3 of 3)****Annex Point IIA, VII, 7.2**

Special investigation in ants

3.3.1	Method of analysis for reference substance	Reference substance not used.
3.4	Testing procedure	
3.4.1	Dilution water	Refer to table A7_1_1_2-1 for description of test site.
3.4.2	Test organisms	Mangrove ant <i>Polyrhachis sokolova</i>
3.4.3	Test system	Refer to table A7_1_1_2-1, at the end of this study summary for description of test site. Nest concentrations of carbon dioxide were determined using air probes at different depths of the nest of the mangrove ant.
3.4.4	Test conditions	The nests of the mangrove ant <i>Polyrhachis sokolova</i> are found in soil in intertidal mangrove communities, and thus are covered in water at high tide for several hours. Some of the nest galleries are flooded, but others retain air pockets where the ants retreat. During and following inundation (where the ants nests were covered in water), carbon dioxide concentrations in the ants nest were measured from air samples collected from different levels in the nest, and from artificial control "holes" in the mud away from the nest. This study measured normal, natural background levels of carbon dioxide in an ant's nest. Refer to table A7_1_1_2-1, at the end of this study summary for description of test site.
3.4.5	Duration of test	11 days.
3.4.6	Test parameter	Carbon dioxide concentration in the ants nest.
3.4.7	Sampling	Gas samples were drawn from the nest chambers while the nest was under water (0 day), 1h after the water receded and then 1, 2, 5, 8 and 11 days after the nest was covered in water (referred to in study report as "inundation"). Control measurements, taken from artificial holes in the mud away from the ants nest were taken on day 5, 6, 8 and 11 after the mud was covered in water.
3.4.8	Monitoring of TS concentration	Refer to 3.4.7 Sampling (above) for details.
3.4.9	Statistics	A Kruskal-Wallis Test was used to compare carbon dioxide concentrations at different depths and from different times in the tidal cycle (time since nest was covered in water, referred to in study report as "inundation") in the nest chambers and from different times in the artificial (control) holes. A Mann-Whitney <i>U</i> -test was used to compare the carbon dioxide concentrations in the control holes at shallow (<10 cm) and deep (> 10cm) levels. A probability level of 0.05 was used to determine statistical significance.
		4 RESULTS
4.1	Limit Test	Not performed.
4.2	Results test substance	
4.2.1	Initial concentrations of test substance	Nest carbon dioxide concentrations were high (2.5% - 11%) during and immediately after the nest was covered in water (referred to in study report as "inundation"), but the carbon dioxide concentration in the upper regions of the nest fell as soil water levels receded. However, at depths >10 cm below the level soil surface the carbon dioxide concentrations remained relatively high and stable (at approximately 2%) over the 11 day test period (between one high tide and the next).

Section A7.4.1.2

Acute toxicity to invertebrates (3 of 3)

Annex Point IIA, VII, 7.2

Special investigation in ants

4.2.2	Actual concentrations of test substance	Refer to "4.2.1 Initial concentrations of test substance" (above) for details.
4.2.3	Effect data (Immobilisation)	The carbon dioxide concentrations in the nests of the mangrove ant <i>Polyrhachis sokolova</i> during high tides are among the highest recorded for insect nests, suggesting that these ants may have unusual physiological attributes to match the behavioural and ecological challenges associated with living in an intertidal zone.
4.2.4	Concentration/response curve	Not reported.
4.2.5	Other effects	None reported.
4.3	Results of controls	Air samples collected from artificial control "holes" in the mud away from the nest. Refer to table A7.1.1.2-3 at the end of this study summary for details of control measurements.
4.4	Test with reference substance	Not performed.
5.1	Materials and Methods	<p>5 APPLICANT'S SUMMARY AND CONCLUSION</p> <p>This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.</p> <p>For details of study sites, see A7_1_1_2-1 at the end of this study summary.</p> <p><u>Determination of nest concentrations of carbon dioxide</u></p> <p>At site 1 (Rapid Creek) 10 air probes were placed in the two nests so that air samples could be extracted from different depths of the nest. The probes consisted of a 30 cm long plastic pipette (1 mL) with an injection needle glued into the upper end, so that a three-way valve could be placed at the top. All probes were placed with their tip into a nest chamber and a layer of plaster of paris was placed on the soil surface around the probe to prevent water intrusion. Gas samples were drawn from the nest chambers while the nest was under water (0 days since inundation), 1h after the water receded and then 1, 2, 5, 8 and 11 days after the water had receded (inundation). Thus, the gaseous environments in the nests were sampled between one high tide and the next (when the nest was covered in water). In addition to the permanent probes at site 1, a portable probe was used at site 2 (East Arm, Darwin Harbour) to take gas samples from the nest when it was under water (during inundation). The portable probe was inserted into the mud until it struck an ant gallery, then the sample was drawn.</p> <p><u>Control samples: Determination of carbon dioxide in soil</u></p> <p>To determine, independently from the ants, the effect of depth in the soil and time since the last high tide when the mud was covered with water (inundation) on carbon dioxide concentrations, six artificial control holes with a diameter of 2-3 cm and depths from 7-25 cm were made at site 1 (Rapid Creek). Permanent probes were placed inside the holes, and the top of the holes were sealed with mud and plaster of paris.</p> <p>The samples of air from the nest chambers and the control holes were withdrawn with a 5 ml glass syringe. By using a second syringe and the three way valve, the probe and the "dead volume" in the syringe was flushed with air from the nest before the sample</p> <p>(continued...)</p>

5.1 Materials and Methods

(Continued)

was taken. The tubes were flushed twice with 2 mL of air and then an air sample of 4-5 mL was drawn. The syringe with the closed three-way valve was removed from the probe and a new three-way valve was placed on the probe.

Sources of carbon dioxide in the nest: Contribution of Ant Respiration to Carbon Dioxide Concentration in Nest

To estimate the contribution of ant respiration to the carbon dioxide concentration of the nests, carbon dioxide production of individual ants and number of ants living in the nest was measured. The carbon dioxide production of the brood was measured at 25°C after placing two broods in the experimental chamber at a time (n= 5 sets of larvae, n = 2 sets of pupae). The respiration chambers were cylindrical glass tubes with a length of 60 mm and a diameter of 13 mm with both ends stopped by rubber plugs. The chambers were placed in a temperature regulated water baths during the measurements. The number of workers was estimated for the two nests at site 1 using a mark-recapture technique. The workers were captured using a small portable vacuum cleaner and trapped in a bag made of nylon mesh which allowed marking of their gaster with white correction ink directly through the mesh without anaesthetising the ants. The ants were captured when leaving the nest entrances, marked and released within 30 min. Ants were recaptured the following day when workers were leaving the nest.

Sources of carbon dioxide in the nest: Contribution of Microbial Respiration in the mud

To assess the contribution of microbial respiration in the mud, independently of the ants, respiration was measured from sediment (mud) samples taken during the construction of the control holes at depths of 0, 2, 7 and 20 cm, and from nests at depths of 5 and 15 cm. The samples were taken horizontally in the holes with a 6 mL plastic syringe (with the tapered end cut off), and immediately sealed with an airtight plastic cap. In the laboratory, the first 1cm of the soil core was removed and then 1mL was taken and placed in the respiration chamber. The respiratory measurements were taken at 25°C.

Determination of Carbon Dioxide in Samples

The air samples from the nests and the respiratory measurements were analysed for carbon dioxide using a flow-through analyser model LI-6251, connected to a data acquisition and analyser system (Sabel System International, Nevada USA, using Datacan V software). The airflow was kept constant at 150 mL per minute.

Determination of Gallery Volumes in the Nest

At the end of the experimental period, the volume of the galleries in Nest A at site 1 (Rapid Creek) was determined by removing two soil cores (2 cm in diameter and 30 cm long) from the centre of the nest, so that galleries in all depths were exposed to the holes. A measured amount of water was added through two funnels until the water level reached the top of the nest.

5.2 Results and discussionNest concentrations of carbon dioxide

Nest concentrations of carbon dioxide in the deeper parts of the *Polyrhachis sokolova* nest fluctuated between 2 and 3%, and although the mean concentrations were generally higher than other parts of the nest, they were only statistically higher during days 0 and 5 after the nest was covered in water (inundation).

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (3 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in ants	

5.2	Results and discussion	<p>The carbon dioxide concentration in the middle level (<10 cm below the soil surface) was significantly higher than in the nest mound 1h after the water receded, but by the next day there was no significant difference among the depths.</p> <p>In the nest mound, the carbon dioxide concentration at 0 days (when the nest was covered in water) was significantly greater than all the other sample periods. At the middle level (<10 cm below soil surface) the carbon dioxide concentration was higher 1h after the water receded compared to all the other time periods. The carbon dioxide concentrations at the deep levels in the nest were not significantly different across all time periods from day 0 (when the nest was covered in water) to 11 days after the water receded.</p> <p>Air samples from the nest at site 2 (East Arm, Darwin Harbour) showed a very high carbon dioxide concentration when it was covered in water, with values up to 11% at depths below 10 cm and 8.7% at depths between 0 and 10cm. One hour after 3h of being covered in water, the carbon dioxide concentration had decreased to 2.2% and 0.8% at the respective depths.</p> <p>By using a portable probe to sample the various ant galleries when the nest was covered in water, it was found that although some galleries were still full of air, others filled with water and this was the case at all depths in the nest. Thus, the nest was not completely water tight and the air volume available to the ants decreased when the nest was covered in water. The high carbon dioxide concentration in the control holes shows that soil respiration is quite significant. As the tide enroaches upon the nest the water level rises from below and at the same time tidal water infiltrates the nest from above. Therefore, carbon dioxide rich air from the soil capillary space will be replaced by water and some of the air will end up in the galleries and be trapped in the air pockets. Thus, the air in the air pockets will reach very high concentrations of carbon dioxide due to the combined effects of the high density of ants and the input of carbon dioxide from the soil. The carbon dioxide concentrations in the nests were most variable at the deepest level, and at all levels when the nest was covered in water. During inundation, the ants aggregated in galleries that are presumably segregated from other galleries due to the flooding of some parts of the nest and these clumps of ants may account for the variability in carbon dioxide at this time.</p> <p>Soon after the water recedes from the nest mound and the middle of the nest < 10 cm deep), the carbon dioxide concentrations fall. The carbon dioxide concentrations are highest in the middle region of the nest 1h after the water has receded, because there is still water in the soil (even though the mound has already drained), and because the nest has been covered in water for at least an hour longer (compared to time 0). The concentrations in the middle region of the nest at this time are similar to those in the deeper galleries when the nest is covered in water. No deep galleries were sampled 1h or 1 day after the nest was covered in water, because all the deep galleries being sampled were full of water. When the</p> <p>(Continued)</p>
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5.2 Results and discussion

water level falls in the soil, fresh atmospheric air is drawn in to the nest galleries through the four nest openings, diluting the air in the upper parts of the nest. This process presumably continues until the water level falls below the deepest galleries in the nest. The carbon dioxide production from the ants and from the mud continually adds carbon dioxide to the nest. Therefore the resulting carbon dioxide concentration depends on the two opposing processes: input of fresh air as the water level falls, and the production of carbon dioxide by ants and mud.

The main factor in the ventilation of the nests of *Polyrhachis sokolova* is the changing water level inside the nest, and without this flushing of air the carbon dioxide concentration in the nest would reach very high levels due to the intense biological activity in the mud. After exposure to fresh air after the water receded, the upper parts of the nest reached equilibrium by the first day after the nest was covered in water and the carbon dioxide concentration remained relatively stable until the next high tide. The deepest parts of the nest experienced a stable carbon dioxide concentration throughout the tidal cycle presumably because deep galleries were less diluted by fresh air and may influx of air caused by falling water levels was from parts of the nest above, rather than atmospheric air.

The extremely high concentrations of carbon dioxide in the nest at site 2 (East Arm, Darwin Harbour) may be due to the location of this site deep within the tidal zone, resulting in a higher tide and longer period of being covered with water. Also the nest at East Arm, Darwin Harbour had no mound so it did not drain as quickly or as often as the nests in site 1 (Rapid Creek).

Carbon dioxide concentrations in the control samples

For results of measurements taken to determine the carbon dioxide concentration in mangrove mud, refer to table A7.1.1.2-3 at the end of this study summary.

The mean carbon dioxide concentrations in the control holes in the soil near the nests were generally higher at deeper depths (>10 cm), but the difference was only statistically significant 11 days after inundation. The carbon dioxide concentrations did not differ across the time periods in the shallow holes, but in the deeper holes (>10 cm), there was a significant decrease in carbon dioxide concentrations that occurred between days 6 and 8 inundation.

Sources of carbon dioxide

For results of measurements taken to determine the sources of carbon dioxide in the nest e.g. due to respiration of the ants, see table A7_1_1_2-2 at the end of this study summary.

Estimates of population density in ant colonies by capture-release-recapture techniques are always encumbered with sources of error due to the lack of complete mixing of marked individuals in the nest. The volume of nest A at site 1 (Rapid Creek) was estimated as 64 L, but it is acknowledged that some water could have leaked during the filing procedure and, on the other hand, some galleries could still have contained air pockets. The carbon dioxide production from ants increased the total carbon dioxide concentration by $(50 \times 1000/640000) 0.008\%$ per hour. Assuming

(continued...)

5.2	Results and discussion	<p>that the mean diameter of the galleries is 2cm, 1L of galleries has a surface area of 1000 cm² giving a total gallery surface of 64000 cm². The carbon dioxide input from mud is (6.2 x 64000 μL/h) 396 mL/h equivalent to an increase in the total nest concentration of 0.6% per hour. Thus the ants only add approximately 10-15% of the total carbon dioxide to the nest, which is consistent with the high values of carbon dioxide found in the control holes. During inundation the air volume available to the ants decreases and the density of the ants in the air pockets is at it's highest and under these conditions the ants may contribute much more to the increase of carbon dioxide concentration.</p>
5.2.1	EC ₀	Not reported.
5.2.2	EC ₅₀	Not reported.
5.2.3	EC ₁₀₀	Not reported.
5.3	Conclusion	<p>Results to this test show that nest carbon dioxide concentrations were high (2.5% - 11%) during and immediately after the nest was covered in water (referred to in study report as "inundation"), but the carbon dioxide concentration in the upper regions of the nest fell as soil water levels receded. However, at depths >10 cm below the level soil surface the carbon dioxide concentrations remained relatively high and stable (at approximately 2%) over the 11 day test period (between one high tide and the next). Carbon dioxide concentrations in muds around the nest can fluctuate between 0.5% and 2% < 10cm below the surface and between 1.2% and 3.% > 10 cm below the surface.</p> <p>Soil microbial respiration is a major source of carbon dioxide in the nests of <i>Polyrhachis sokolova</i>, but if large numbers of ants are restricted to relatively small portions of the nest (e.g. during inundation), then ant respiration is a mJOR contributor to carbon dioxide concentrations during high tides.</p>
5.3.1	Reliability	3
5.3.2	Deficiencies	<p>Yes.</p> <p>This study was not carried out to EC Method C2 in Annex V of Directive 67/548/EEC.</p> <p>Rather than looking at acute toxicity <i>per se</i>, this report measures the natural fluctuations in background carbon dioxide concentrations in a the nests of the mangrove ant, <i>Polyrhachis sokolova</i>.</p> <p>It should be noted that the use of carbon dioxide by Rentokil Initial would not increase the normal atmospheric concentrations of carbon dioxide in the locality.</p>

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (3 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in ants	

Table A7_1_1_2-1 Description of test site

Criteria	
Description of site	<p>The study was performed in the mangroves around Darwin, NT Australia (12°30'S 131°00'E), in the dry season of 2002.</p> <p><u>Site 1</u> The main experimental site (site 1) was established on the east bank of Rapid Creek, approximately 150m from the creek and 600 m from the sea neighbouring the Charles Darwin University, and included two nests, labelled A and B. The site is very well protected, vegetated by 4-5 m tall trees of <i>Rhizophora stylosa</i> and <i>Avicennia marina</i>.</p> <p><u>Site 2</u> This site was located at East Arm in Darwin Harbour near the former World War II quarantine area. The site is only 100m from the harbour and is quite exposed, so waves can often occur during inundation. The vegetation is a pure stand of <i>Rhizophora stylosa</i> 5-7 m tall.</p>
Description of test nests.	<p><u>General description of mangrove ants nest</u> Nests of <i>Polyrhachis sokolova</i> sometimes include a central mound elevated above the surrounding soil surface. The size and shape of the mounds were measured at the two sites, and elevations were calculated by measuring the height of the water level above the nest at maximum tide, where the height above the lowest astronomical tide (LAT) is tabulated for the Darwin area.</p> <p><u>Site 1</u> The surface is very muddy with many crab holes and piles of excavated mud from the ants' digging activity. The elevation of the nests at this site (Rapid Creek) were 6.75 cm above lowest astronomical tide (LAT) for nest A, and 6.65 cm above LAT for nest B. This means that nest B was inundated during 27% of the high tides with up to 1.35m of seawater. The mound of nest A has a diameter of 100 cm and a height 20-25 cm above the surroundings, whereas the mound of nest B only reached 17 cm and the diameter was 80 cm. Both nests had four entrances.</p> <p><u>Site 2</u> The substrate is sandy on the surface but very muddy below 1-2 cm. There is very little sign of crab activity, so the surface was smooth. The nest at this site (East Arm) was 5.85 cm above lowest astronomical tide (LAT), thus it was inundated by more than 2.15 m of seawater during more than 64% of high tides. The nest had no mound, but there were volcano-like walls around the two entrances 4-5 cm high.</p>

Rentokil Initial plc	Carbon Dioxide	March 2004
Section A7.4.1.2	Acute toxicity to invertebrates (3 of 3)	
Annex Point IIA, VII, 7.2	Special investigation in ants	

Table A7_1_1_2-2 Sources of carbon dioxide in the nests at site 1 (Rapid Creek)

Criteria			
The specific respiratory rates +/- SD for individuals workers of <i>Polyrhachis sokolova</i> *	At 15°C At 25°C At 35°C	1.45 +/-0.02 µL CO2/mg dry weight/h 2.79 +/-0.74 µL CO2/mg dry weight/h 4.27 +/- 0.94 µL CO2/mg dry weight/h	
The specific respiratory rates +/- SD for larvae and pupae of <i>Polyrhachis sokolova</i>	At 25°C	Larvae 0.50 +/- 0.06 µL CO2 /mg dry weight/h Pupae 0.56 +/- 0.04 µL CO2/mg dry weight/h	
The mean (+/- SD) fresh and dry weights for workers (n = 10)		Fresh weight : 17.7 +/- 2.3 mg Dry weight: 5.9 +/- 0.8 mg	
The crude measurements of mud respiration from the nest		23.1 +/- 7.3 µL CO2/mg fresh weight/h (or 6.2 +/- 2.0 µL CO2/ cm ² surface of sample per hour).	
The population estimates of foraging workers at site 1		1051 +/-166 individuals in nest A 2065 +/- 463 individuals in nest B.	
The measured volume of galleries in nest A		64 L.	

Footnote: The population estimates apply only to the foraging population which can differ strongly from the total population. No information is available on the foraging behaviour of *Polyrhachis sokolova* so, for the calculations cited below, it is assumed that half the population is foraging. The total respiratory rate at 25°C by workers in nest A (site 1 Rapid Creek) can be calculated as (2 x 1050 x 5.9 x 2.8 µL/h) 34.7 mL/h. Assuming that the brood respiration is half the worker population the total carbon dioxide production in the nest would be approximately 50 mL h⁻¹ at 25°C and approximately 100 mL h⁻¹ at 35°C.

Table A7_1_1_2-3 Carbon dioxide concentration in mangrove mud (artificial holes, acting as control measurements)

Days since mud was covered with water (inundation)	Concentration of carbon dioxide measured in mud	
	Shallow hole (< 10 cm)	Deep hole (> 10cm)
5	ca. 2.1%	3.5 %
6	ca. 2.3%	3.3%
8	0.5%	1.4%
11	0.5%	1.2%

Evaluation by Competent Authorities	
	Use separate "evaluation boxes" to provide transparency as to the comments and views submitted.
	EVALUATION BY RAPPORTEUR MEMBER STATE
Date	<i>Give date of action</i>
Materials and Methods	<i>State if applicants version is acceptable, or indicate relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion.</i>
Results and discussion	<i>Adopt applicant's version or include revised version. If necessary, discuss relevant deviations from applicant's view referring to the (sub)heading numbers.</i>
Conclusion	Other conclusions: <i>(adopt applicant's version or include revised version)</i>
Reliability	<i>Based on assessment of materials and methods include appropriate reliability indicator.</i>
Acceptability	acceptable / not acceptable <i>(give reasons if necessary e.g. if a study is considered acceptable despite a poor reliability indicator. Discuss the relevance of deficiencies and indicate if repeat if necessary).</i>
Remarks	
	COMMENTS FROM
Date	<i>Give date of comments submitted.</i>
Materials and Methods	<i>Discuss additional relevant discrepancies referring to the (sub)heading numbers and to applicant's summary and conclusion</i> <i>Discuss if deviating from view of rapporteur member state. .</i>
Results and discussion	<i>Discuss if deviating from view of rapporteur member state.</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state.</i>
Reliability	<i>Discuss if deviating from view of rapporteur member state.</i>
Acceptability	<i>Discuss if deviating from view of rapporteur member state.</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.1.3 Annex Point/TNsG Annex IIA, VII.7.3	Growth inhibition test on algae Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input type="checkbox"/>	Technically not feasible <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification <input type="checkbox"/>
Detailed justification:		It is not scientifically necessary to calculate the growth inhibition of CO ₂ to algae because CO ₂ is an essential substrate for photosynthesis. In addition, under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water, outside normal atmospheric ranges. <i>See next page for full details of the scientific calculation, which supports this statement.</i> Continued.....



Table 4-2: Standard form for justification of the non-submission of data

<p>Section 7.4.1.4 Annex Point/TNsG Annex IIA, VII.7.4, Annex IIIA, VII.3</p>	<p>Inhibition to microbiological activity Section 7: Ecotoxicological Profile, including Fate and Behaviour</p>	
<p>JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		<p>Official use only</p>
<p>Other existing data</p>	<p><input checked="" type="checkbox"/> Technically not feasible []</p>	<p>Scientifically unjustified []</p>
<p>Limited exposure</p>	<p><input checked="" type="checkbox"/> Other justification []</p>	
<p>Detailed justification:</p>	<p>Whilst elevated levels of dissolved CO₂ may affect environmental conditions for bacteria by reducing pH, there are a number of mitigating factors that would reduce any environmental impacts of such changes and which make it unnecessary to generate new test data.</p> <p>a) Most free-living prokaryotic bacteria can tolerate a pH range of about 3 units (three orders of magnitude changes in H⁺ ¹</p> <p>b) There is a high level of functional redundancy amongst mixed communities of micro-organisms such that declines in population of some species e.g. due to unfavourable pH conditions, will be compensated for by increases in others. The effect of this biological diversity and different environmental optima for different species means that bacteria can live in a wide range of pH conditions, from 0.5 – 9.0. ¹</p> <p>In addition, it is not scientifically necessary to determine the effect of increased carbon dioxide on microbial activity, because under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water, outside normal atmospheric ranges.</p> <p><i>See next page for full details of the scientific calculation, which supports this statement.</i></p> <div style="background-color: black; width: 100%; height: 60px; margin-top: 10px;"></div> <p>Continued.....</p>	



Section 7.4.1.4 Annex Point/TNsG Annex IIIA, VII.7.4, Annex IIIA, VII.3	Inhibition to microbiological activity Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:
(continued):

[REDACTED]

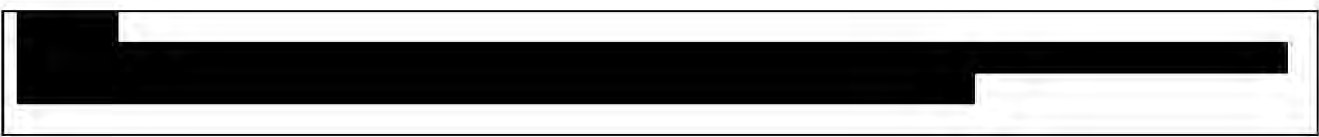
Undertaking of intended data submission [] Not applicable

Evaluation by Competent Authorities	
	Use separate "evaluation boxes" to provide transparency as to the comments and views submitted
	EVALUATION BY RAPPORTEUR MEMBER STATE
Date	<i>Give date of action</i>
Evaluation of applicant's justification	<i>Discuss applicant's justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant's justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
	COMMENTS FROM OTHER MEMBER STATES (specify)
Date	<i>Give date of comments submitted</i>
Evaluation of applicant's justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>

Remarks

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.2 Annex Point/TNsG Annex IIA, VII.7.5	Bioconcentration Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input checked="" type="checkbox"/>
Limited exposure	<input type="checkbox"/>	Other justification	<input type="checkbox"/>		
Detailed justification:	<p>“Bioconcentration” is the process leading to a higher concentration of, for example, a pesticide in an organism than in environmental media to which it is exposed.</p> <p>Since CO₂ is a naturally occurring substance that all living organisms are exposed to, and which plays a vital role in the normal maintenance of life, studies into the bioconcentration are not justified. The partition coefficient of CO₂ is 2.26 (isobutanol/water) and 1.74 (olive oil/water). The n-octanol/water partition coefficient has been calculated to be 0.83.</p>				
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable			



Section 7.4.2 Annex Point/TNsG Annex IIA, VII.7.5	Bioconcentration Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
	Use separate “evaluation boxes” to provide transparency as to the comments and views submitted
	EVALUATION BY RAPPORTEUR MEMBER STATE
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
	COMMENTS FROM OTHER MEMBER STATES (specify)
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3 Annex Point/TNsG Annex IIIA, XIII.2	Effects on Aquatic Organisms, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input checked="" type="checkbox"/>		
Detailed justification:	<p>Effects on aquatic organisms is not required as carbon dioxide is not intended to be either used or released into aquatic environments.</p> <p>For these purposes, it is intended that CO₂ be used as a biocide in a closed system.</p> <p>These studies are only required if the results of 7.4.1.1, 7.4.1.2, 7.4.1.3 and 7.4.1.4 indicate a danger to the environment, as this is not indicated further tests are not required.</p>				
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.			



Section 7.4.3 Annex Point/TNsG Annex IIIA, XIII.2	Effects on Aquatic Organisms, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.1 Annex Point/TNsG Annex IIIA, XIII.2.1	Prolonged toxicity to an appropriate species of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input type="checkbox"/>	Technically not feasible <input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Scientifically unjustified <input type="checkbox"/>
Detailed justification:	<p>This test is not usually required. Results of 7.4.1.1, 7.4.1.2, 7.4.1.3 and 7.4.1.4 do not indicate further testing is required.</p> <p>Carbon dioxide is not intended to be either used or released into aquatic environments.</p> <p>For these purposes, it is intended that CO₂ be used as a biocide in a closed system.</p>	
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.



Section 7.4.3.1 Annex Point/TNsG Annex IIIA, XIII.2.1	Prolonged toxicity to an appropriate species of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FROM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.2 Annex Point/TNsG Annex IIIA, XIII.2.2	Effects on reproduction and growth rate of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input checked="" type="checkbox"/>		
Detailed justification:	<p>These data are only required if the data given in the core base set indicates a need for further testing. A requirement for further testing is not indicated.</p> <p>It is not necessary because under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water, outside normal atmospheric ranges. Study summaries are given in the core base data set for two pieces of work investigating the effects of carbon dioxide to fish. One of which shows that dissolved concentrations of up to 6.3% carbon dioxide have not given rise to irreversible physiological and behavioural effects. The core base data set also shows that substantial rises in atmospheric carbon dioxide are never going to occur under the normal conditions of use of Rentokil Initial's biocidal products and the data cited in this application, it is not necessary to conduct further studies.</p>				
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.			



Section 7.4.3.2 Annex Point/TNsG Annex IIIA, XIII.2.2	Effects on reproduction and growth rate of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.3.1 Annex Point/TNsG Annex IIIA, XIII.2.3	Bio-accumulation in an appropriate species of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour				
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input checked="" type="checkbox"/>		
Detailed justification:	This information is only required when there is a risk of secondary poisoning or there are other features indicating bio-accumulation. There is no risk of secondary poisoning of fish with the use of carbon dioxide. Carbon dioxide is a naturally occurring substance that all living organisms are exposed to, and which plays a vital role in the normal maintenance of life. Carbon dioxide in this case is used in a closed system, it is later released to the atmosphere and is not available to the aquatic environment.				
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.			



Section 7.4.3.3.1 Annex Point/TNsG Annex IIIA, XIII.2.3	Bio-accumulation in an appropriate species of fish. Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.3.2 Annex Point/TNsG Annex IIIA, XIII.2.4	Bio-accumulation in an appropriate invertebrate species. Section 7: Ecotoxicological Profile, including Fate and Behaviour				
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input checked="" type="checkbox"/>		
Detailed justification:		This information is only required when there is a risk of secondary poisoning or there are other features indicating bio-accumulation. There is no risk of secondary poisoning of fish with the use of carbon dioxide. Carbon dioxide is a naturally occurring substance that all living organisms are exposed to, and which plays a vital role in the normal maintenance of life. Carbon dioxide in this case is used in a closed system, it is later released to the atmosphere and is not available to the aquatic environment.			
Undertaking of intended data submission	<input type="checkbox"/>	Not applicable.			



Section 7.4.3.3.2 Annex Point/TNsG Annex IIIA, XIII.2.4	Bio-accumulation in an appropriate invertebrate species. Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.4 Annex Point/TNsG Annex IIIA, XIII.2.4	Effects on reproduction and growth rate with an appropriate invertebrate species. Section 7: Ecotoxicological Profile, including Fate and Behaviour	
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only
Other existing data <input type="checkbox"/>	Technically not feasible <input type="checkbox"/>	Scientifically unjustified <input type="checkbox"/>
Limited exposure <input checked="" type="checkbox"/>	Other justification <input checked="" type="checkbox"/>	
Detailed justification:	This information is only required when chronic exposure is expected or there are other features indicating the need for this test. There will be no chronic exposure as described in the core base data set the carbon dioxide is a naturally occurring substance that all living organisms are exposed to, and which plays a vital role in the normal maintenance of life. Carbon dioxide in this case is used in a closed system, it is later released to the atmosphere and is not available to the aquatic environment.	
Undertaking of intended data submission <input type="checkbox"/>	Not applicable.	



Section 7.4.3.4 Annex Point/TNsG Annex IIIA, XIII.2.4	Effects on reproduction and growth rate with an appropriate invertebrate species. Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.5 Annex Point/TNsG Annex IIIA, XIII.3.4	Effects on any other specific, non-target organisms (flora and fauna) believed to be at risk. Section 7: Ecotoxicological Profile, including Fate and Behaviour	
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only
Other existing data <input type="checkbox"/>	Technically not feasible <input type="checkbox"/>	Scientifically unjustified <input type="checkbox"/>
Limited exposure <input checked="" type="checkbox"/>	Other justification <input checked="" type="checkbox"/>	
Detailed justification:	<p>This information is only required if the data from other ecotoxicity tests indicates the need to do so, or if there is a need indicated by the intended use.</p> <p>Other ecotoxicity tests on carbon dioxide do not indicate that further testing is required.</p> <p>In addition, It is not scientifically necessary to determine the effect of increased carbon dioxide on non-target flora and fauna because under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water or air, outside normal atmospheric ranges. <i>See next page for full details of the scientific calculation, which supports this statement.</i></p> <p align="center">(Continued...)</p>	

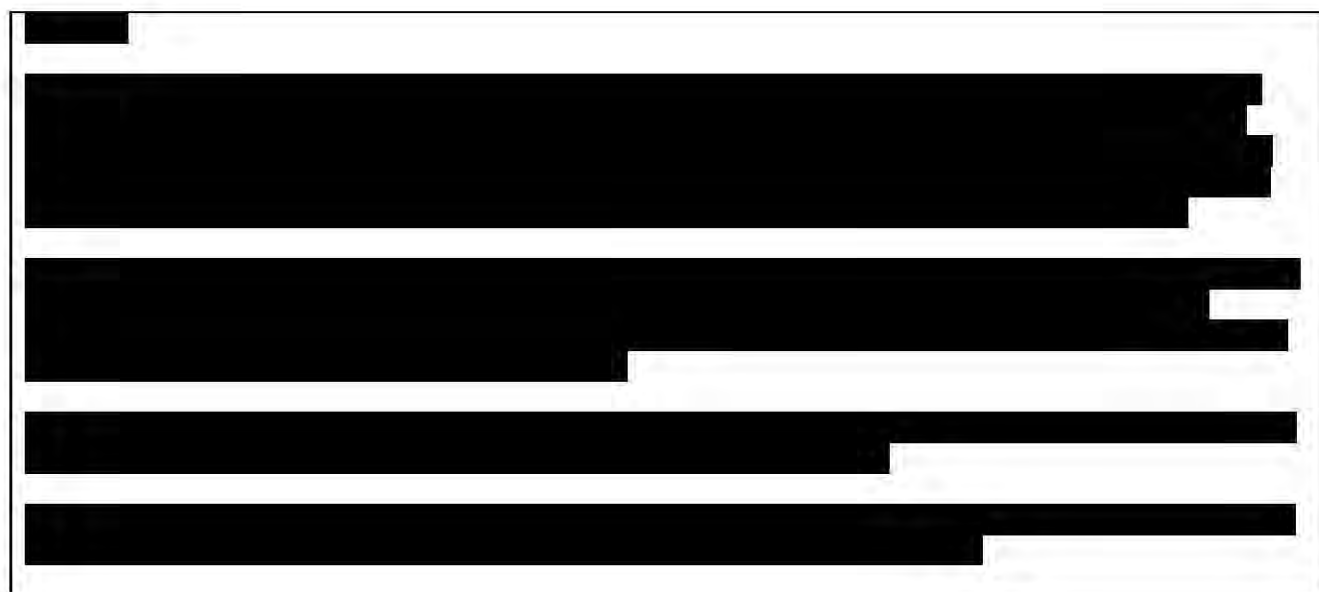


Section 7.3.2 Annex Point/TNsG Annex IIIA, XII.3	Fate and behaviour in air, further studies Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FROM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.5.1 Annex Point/TNsG Annex IIIA, XIII.3.4	Effects on sediment dwelling organisms Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input type="checkbox"/>		
<p>Detailed justification:</p> <p>This information is only required if the active substance partitions to, and persists in, aquatic sediments such that sediment dwelling organisms are likely to be exposed to the active substance.</p> <p>Under normal conditions of use, the use of carbon dioxide in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water (outside normal atmospheric ranges) thus sediment dwelling organisms will not be exposed to increased carbon dioxide. This makes it unnecessary to generate data on the effects of increased carbon dioxide to sediment dwelling organisms.</p> <p><i>See next page for full details of scientific calculation, which supports this statement.</i></p> <p>(Continued.....)</p>					



Section 7.4.3.5.1 Annex Point/TNsG Annex IIIA, XIII.3.4	Effects on sediment dwelling organisms Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:

[REDACTED]

[REDACTED]

[REDACTED]

Undertaking of intended data submission Not applicable. []

Section 7.4.3.5.1 Annex Point/TNsG Annex IIIA, XIII.3.4	Effects on sediment dwelling organisms Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FORM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.4.3.5.2 Annex Point/TNsG Annex IIIA, XIII.3.4	Aquatic plant toxicity Section 7: Ecotoxicological Profile, including Fate and Behaviour		Official use only
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input checked="" type="checkbox"/>
Detailed justification:	<p>Under normal conditions of use, the use of carbon dioxide by Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the level of carbon dioxide in water (outside normal atmospheric ranges) thus aquatic plants will not be exposed to increased carbon dioxide. This makes it unnecessary to generate data on the effects on increased carbon dioxide to aquatic plants. <i>See next page for full details of scientific calculation which supports this statement.</i></p> <p>Notwithstanding the above, it should be noted that carbon dioxide plays a vital role in the photosynthesis pathway of plants. It is widely accepted that commercial horticulturists use carbon dioxide to enrich the atmospheres of their greenhouses and other growing environments to accelerate the growth of crops.</p> <p>(Continued...)</p>		



Section 7.4.3.5.2 Annex Point/TNsG Annex IIIA, XIII.3.4	Aquatic plant toxicity Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:

[REDACTED]

[REDACTED]

[REDACTED]

Undertaking of intended data submission Not applicable. []

Section 7.4.3.5.2 Annex Point/TNsG Annex IIIA, XIII.3.4	Aquatic plant toxicity Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FROM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Table 4-2: Standard form for justification of the non-submission of data

Section 7.5.1.1 Annex Point/TNsG Annex IIA, VII.7.4	Inhibition to microbial activity (terrestrial) Section 7: Ecotoxicological Profile, including Fate and Behaviour				
<p align="center">JUSTIFICATION FOR NON-SUBMISSION OF DATA</p> <p><i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i></p> <p><i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i></p>		Official use only			
Other existing data	<input type="checkbox"/>	Technically not feasible	<input type="checkbox"/>	Scientifically unjustified	<input type="checkbox"/>
Limited exposure	<input checked="" type="checkbox"/>	Other justification	<input type="checkbox"/>		
Detailed justification:	<p>This information is only required if a concern for the terrestrial compartment is indicated by the risk assessment or if there is likely to be long term exposure to the active substance. Carbon dioxide, under normal conditions of use in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the levels of carbon dioxide found in terrestrial systems, outside normal atmospheric ranges.</p> <p><i>Refer to next page for details of scientific calculation which supports this statement.</i></p> <p>In addition, there is no mechanism for the carbon dioxide to be released directly into terrestrial systems. Consequently, there will be no increased levels of carbon dioxide in terrestrial systems making it unnecessary to determine the effect of increased carbon dioxide on microbial activity.</p> <p>Continued.....</p>				



Section 7.5.1.1 Annex Point/TNsG Annex IIA, VII.7.4	Inhibition to microbial activity (terrestrial) Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Detailed justification:
 (Continued)

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

[REDACTED]

Undertaking of intended data submission Not applicable

Section 7.5.1.1 Annex Point/TNsG Annex II A, VII.7.4	Inhibition to microbiological activity (terrestrial) Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate “evaluation boxes” to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant’s justification	<i>Discuss applicant’s justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant’s justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FROM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant’s justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Section A7.5.1.2 Annex Point / TNsG Annex IIIA XIII 3.2	Earthworm, acute toxicity test Section 7: Ecotoxicological Profile, including Fate and Behaviour	
JUSTIFICATION FOR NON-SUBMISSION OF DATA <i>As outlined in the TNsG on data requirements, the applicant must always be able to justify the suggested exemptions from the data requirements. The justifications are to be included in the respective location (section) of the dossier.</i> <i>If one of the following reasons is marked, detailed justification has to be given below. General arguments are not acceptable</i>		Official use only
Other existing data	<input checked="" type="checkbox"/> Technically not feasible []	<input type="checkbox"/> Scientifically unjustified []
Limited exposure	<input checked="" type="checkbox"/> Other justification []	
Detailed justification:	<p>This data is only required if a concern for the terrestrial compartment is indicated by the risk assessment or if there is likely to be long-term exposure to the active substance.</p> <p>Carbon dioxide, under normal conditions of use in Rentokil Initial's rodenticide (PT14) products will not cause any elevation in the levels of carbon dioxide naturally found in terrestrial systems, outside normal atmospheric ranges. In addition, there is no mechanism for the carbon dioxide to be released directly into the terrestrial system. <i>Refer to next page for details of scientific calculation which supports this statement.</i></p> <p>Consequently, there will be no increased carbon dioxide levels in the terrestrial system, so it is not necessary to determine the effect of increased carbon dioxide on earthworms.</p> <p>Notwithstanding this, there is a study available in the public domain which gives an indication about the possible effects increased CO₂ may have on cast production by earthworms. This data has been summarised here, for information.</p> <p>Continued.....</p>	





Section A7.5.1.2 Annex Point / TNsG Annex IIIA XIII 3.2	Earthworm, acute toxicity test Section 7: Ecotoxicological Profile, including Fate and Behaviour
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Evaluation by Competent Authorities	
Use separate "evaluation boxes" to provide transparency as to the comments and views submitted	
EVALUATION BY RAPPORTEUR MEMBER STATE	
Date	<i>Give date of action</i>
Evaluation of applicant's justification	<i>Discuss applicant's justification and, if applicable, deviating view</i>
Conclusion	<i>Indicate whether applicant's justification is acceptable or not. If unacceptable because of the reasons discussed above, indicate which action will be required, e.g. submission of specific test/study data</i>
Remarks	
COMMENTS FROM OTHER MEMBER STATES (specify)	
Date	<i>Give date of comments submitted</i>
Evaluation of applicant's justification	<i>Discuss if deviating from view of rapporteur member state</i>
Conclusion	<i>Discuss if deviating from view of rapporteur member state</i>
Remarks	

Section A7.5.1.2
Annex Point IIIA XIII 3.2

Earthworm, acute toxicity test

		1 REFERENCE	Official use only
1.1	Reference	[REDACTED]	
1.2	Data protection	[REDACTED]	
1.2.1	Data owner	[REDACTED]	
1.2.3	Criteria for data protection	[REDACTED]	
		2 GUIDELINES AND QUALITY ASSURANCE	
2.1	Guideline study	No Not carried out to OECD Guideline 207.	
2.2	GLP	No data	
	<i>(only where required)</i>	GLP was not mentioned in publication.	
2.3	Deviations	Yes This study was not carried out to look at earthworm toxicity <i>per se</i> thus did not follow the OECD test guideline 207 (Earthworm acute toxicity tests). This report investigates the activity levels of earthworms exposed to elevated carbon dioxide levels.	
		3 MATERIALS AND METHODS	
3.1	Test material	As given in section 2.	
3.1.1	Lot/Batch number	[REDACTED]	
3.1.2	Specification	[REDACTED]	
3.1.3	Purity	[REDACTED]	

Section A7.5.1.2 Earthworm, acute toxicity test
Annex Point IIIA XIII 3.2

3.1.4	Composition of product	Not applicable for active substance.	
3.1.5	Further relevant properties	Not reported.	
3.1.6	Method of analysis	The number of surface casts produced was measured every nine days for a year.	
3.2	Reference substance	No	
3.2.1	Method of analysis for reference substance	Not applicable as reference substance was not used.	
3.3	Testing procedure		
3.3.1	Preparation of the test substance.	Not applicable. Carbon dioxide is not pre-treated.	
3.3.2	Application of the test substance	Carbon dioxide added to soil plots using a screen aided CO ₂ control facility (SACC, open top and bottom rings).	
3.3.3	Test organisms	Natural population of surface casting earthworms. Species not given.	
3.3.4	Test systems	Artificial soil test substrate	Not relevant as natural soil used: calcareous grassland classified as transition Rendzina, pH 6.5, bulk density on the top soil 1.1g/cm ³ well developed, stone free loamy topsoil and a rapid transition at 10 –15cm depth to underlying scree. Depth of Ah-horizon averaged 15cm.
		Test mixture	Natural soil compared to the same soil with carbon dioxide added.
		Size, volume and material of test container	Not given in this report but in reference by Leadley PW and Korner Ch (1996) in Korner Ch, Bazzaz FA (eds) Carbon dioxide, populations and communities, Academic Press, San Diego pp 159 -175
		Amount of artificial soil (kg)/ container	Not given in this report but in reference by Leadley PW and Korner Ch (1996) in Korner Ch, Bazzaz FA (eds) Carbon dioxide, populations and communities, Academic Press, San Diego pp 159 -175
		Nominal levels of test concentrations	Control 350µl CO ₂ /l Test 610µl CO ₂ /l
		Number of replicates/concentration	8
		Number of earthworms /test concentration	Not given but natural population
		Number of earthworms/container	Not given but natural population
		Light source	Natural

Section A7.5.1.2 Earthworm, acute toxicity test
Annex Point IIIA XIII 3.2

	Test performed in closed vessels due to significant volatility of test substrate	No. Screen – aided control facility, open top and open bottom rings.
3.3.5 Test conditions	Test temperature	Mean annual air temperature 8.7°C
	Moisture content	Mean annual soil water content was 10% greater ($P<0.01$) in elevated CO ₂ plots (33% dry mass) than in ambient CO ₂ plots (30% dry mass). Mean soil water content measured in treated plots ($26.8 \pm 0.6\%$ dry mass) over the 6 week long summer dry period in 1995 was significantly ($P<0.05$) greater than that measured in the control plots over the same period ($23.3 \pm 0.8\%$ dry mass). When averaged over the entire 13- week dry (6 weeks) and subsequent wetter period (7 weeks), soil water content in treated plots was $29.7 \pm 0.7\%$ and that in untreated plots was $27.0 \pm 0.8\%$ ($p<0.05$).
	pH	6.5 in soil, no data given throughout experiment.
	Adjustment of pH	No
	Light intensity / photo-period	Natural light
	Relevant degradation products.	No degradation products.
3.3.6 Test duration	March 1994 to April 1996. 2 years	
3.3.7 Test parameter	Cast production	
3.3.8 Examination	Each 9 days	
3.3.9 Monitoring of test substance concentration	Continuous control except for a period 15 December 1995 to 6 March 1996 when air temperature below freezing or ground covered in snow.	
3.3.10 Statistics	Data analysed using repeated measurement ANOVA's (Sokal and Rohlf 1981 Biometry. The principles and practice of statistics in biological research, 2 nd edn., Freeman, New York and SYSTAT (1992) Statistics, version 5.2 Systat, Evanston.	
	4. RESULTS	
4.1 Filter paper test	Not performed.	
4.1.1 Concentration	Not applicable as filter paper test not performed.	
4.1.2 Number/percentage of animals showing adverse effects	Not applicable as filter paper test not performed.	

Section A7.5.1.2 Earthworm, acute toxicity test
Annex Point IIIA XIII 3.2

4.1.3 Nature of adverse effects Not applicable as filter paper test not performed,

4.2 Soil Test

4.2.1 Initial concentration of test substance Treatment maintained at 610 µl CO₂/l.

4.2.2 Effect data (Mortality) Mortality not measured

4.2.3 Concentration / response curve Mortality not mentioned

4.2.4 Other effects Rates of surface cast production expressed as g/m/day in communities with elevated CO₂ were up to 6 times higher than those in ambient CO₂.
 Cumulative surface cast production after 1 year was 35% greater in communities with elevated CO₂ than those in ambient CO₂.
 CO₂ induced stimulation of earthworms increased soil turnover.
 CO₂ induced stimulation of earthworms increased N and C cycling.

4.3 Results of controls

4.3.1 Mortality Not reported

4.3.2 Number/percentage of earthworms showing adverse effects Not reported

4.3.3 Nature of adverse effects Not applicable

4.4 Test with reference substance Not performed

4.4.1 Concentration Not required

4.4.2 Results Not required

5 APPLICANT'S SUMMARY AND CONCLUSION

5.1 Materials and methods This study was not carried out to Guideline C.8 in Annex V of Directive 67/548/EEC or to to OECD Guideline 207. Rather than looking at acute toxicity *per se*, this report investigates the effects on cast production of an increased concentration of carbon dioxide.

Using a screen aided CO₂ control facility 1.2m³ plots of grassland on a south west facing slope at an elevation of around 515m near a village of Nenzlingen in the Jura Mountains of Switzerland were set up. Eight were maintained at ambient CO₂ concentrations (350µl CO₂/l) and eight at elevated CO₂ (610µl CO₂/l). Cumulative earthworm surface cast production was measured 40 times over 1 year. Precipitation and soil temperature were continuously monitored. Vegetation from each plot was harvested down to 5cm height in June and October and dried at 80°C for 48 hours, plant dry mass was then calculated. Total C and N content of