

Helsinki, 24 June 2021

**Addressees**

Registrants of JS\_Monoazored\_PR3 as listed in the last Appendix of this decision

**Date of submission of the dossier subject to this decision**

23 May 2019

**Registered substance subject to this decision ("the Substance")**

Substance name: 1-(4-methyl-2-nitrophenylazo)-2-naphthol

EC number: 219-372-2

CAS number: 2425-85-6

**Decision number:** Please refer to the REACH-IT message which delivered this communication (in format CCH-D-XXXXXXXXXX-XX-XX/F)**DECISION ON A COMPLIANCE CHECK**

Under Article 41 of Regulation (EC) No 1907/2006 (REACH), you must submit the information listed in A.1 and A.2 below by **31 March 2022** and all other information listed below by **1 June 2023**.

Requested information must be generated using the Substance unless otherwise specified.

We note that the Substance has been notified as a nanoform under the French nanoparticulate substances reporting system.<sup>1</sup> This indicates that the Substance is manufactured or imported in the European Union in nanoforms, possibly by any addressee of the present decision. However, the REACH Regulation (as amended by Regulation Commission Regulation (EU) 2018/1881) sets out explicit information requirements for nanoforms of substances. Manufacturers and importers of nanoforms must have fulfilled these specific information requirements by 1<sup>st</sup> January 2020. As far as the registration dossier currently submitted on the Substance does not cover any nanoform, the incompliances identified in the present decision relate only to information required on non-nanoforms.

Based on the above, the requested information must be generated using exclusively non-nanoforms of the Substance.

**A. Information required from all the Registrants subject to Annex VII of REACH**

1. Water solubility (Annex VII, Section 7.7.; test method: EU A.6./OECD TG 105)
2. Partition coefficient n-octanol/water (Annex VII, Section 7.8.; test method: EU A.8 or OECD TG 117 or OECD TG 123)
3. Same *In vitro* cytogenicity study in mammalian cells (test method: OECD TG 473) or *In vitro* micronucleus study (test method: OECD TG 487), as request B.1
4. Same *in vivo* genotoxicity study, as request B.2.

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<sup>1</sup> «Dispositif de déclaration des substances à l'état nanoparticulaire », Decree 2012-232 of French Conseil d'Etat of 17 february 2012.

**B. Information required from all the Registrants subject to Annex VIII of REACH**

1. In vitro cytogenicity study in mammalian cells (Annex VIII, Section 8.4.2.; test method: OECD TG 473) or In vitro micronucleus study (Annex VIII, Section 8.4.2.; test method: OECD TG 487)
2. *In vivo* genotoxicity study to be selected according to the following scenarios:
  - a. If the **test results of the *in vitro* study requested in B.1** (*in vitro* cytogenicity study in mammalian cells) are **negative**:

*In vivo* mammalian alkaline comet assay (test method: OECD TG 489) in rats, oral route, on the tissues: liver, glandular stomach and duodenum

OR

Transgenic rodent somatic and germ cell gene mutation assay (test method: OECD TG 488 from 2020<sup>2</sup>) in transgenic mice or rats, oral route on the following tissues: liver oral: and glandular stomach; duodenum must be harvested and stored for up to 5 years. The duodenum must be analysed if the results of the glandular stomach and of the liver are negative or inconclusive

- b. If the **test results of *in vitro* study requested in B.1** (*in vitro* cytogenicity study in mammalian cells) are **positive**:

*In vivo* mammalian alkaline comet assay (test method OECD TG 489) combined with *in vivo* mammalian erythrocyte micronucleus test (test method: OECD TG 474) in rats, oral route. For the comet assay the following tissues shall be analysed: liver, glandular stomach and duodenum.

Reasons for the requests are explained in the following Appendices entitled "Reasons to request information required under Annexes VII to VIII of REACH", respectively.

**Information required depends on your tonnage band**

You must provide the information listed above for all REACH Annexes applicable to you, and in accordance with Articles 10(a) and 12(1) of REACH:

- the information specified in Annex VII to REACH, for registration at 1-10 tonnes per year (tpa), or as a transported isolated intermediate in quantity above 1000 tpa;
- the information specified in Annexes VII and VIII to REACH, for registration at 10-100 tpa.

You are only required to share the costs of information that you must submit to fulfil your information requirements.

For certain endpoints, ECHA requests the same study from registrants at different tonnages. In such cases, only the reasoning why the information is required at lower tonnages is provided in the corresponding Appendices. For the tonnage where the study is a standard information requirement, the full reasoning for the request including study design is given. Only one study is to be conducted; the registrants concerned must make every effort to reach an agreement as to who is to carry out the study on behalf of the other registrants under

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<sup>2</sup> The updated OECD TG 488, adopted on 26 June 2020, is available on OECD website at <https://www.oecd-ilibrary.org/docserver/9789264203907-en.pdf?expires=1596539942&id=id&accname=guest&checksum=D552783C4CB0FC8045D04C88EFFBFA66>.

Article 53 of REACH.

### **How to comply with your information requirements**

To comply with your information requirements you must submit the information requested by this decision in an updated registration dossier by the deadline indicated above. You must also update the chemical safety report, where relevant, including any changes to classification and labelling, based on the newly generated information.

You must follow the general testing and reporting requirements provided under the Appendix entitled "Requirements to fulfil when conducting and reporting new tests for REACH purposes". In addition, you should follow the general recommendations provided under the Appendix entitled "General recommendations when conducting and reporting new tests for REACH purposes". For references used in this decision, please consult the Appendix entitled "List of references".

### **Appeal**

This decision, when adopted under Article 51 of REACH, may be appealed to the Board of Appeal of ECHA within three months of its notification to you. Please refer to <http://echa.europa.eu/regulations/appeals> for further information.

### **Failure to comply**

If you do not comply with the information required by this decision by the deadline indicated above, ECHA will notify the enforcement authorities of your Member State.

Authorised<sup>3</sup> under the authority of Christel Schilliger-Musset, Director of Hazard Assessment

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<sup>3</sup> As this is an electronic document, it is not physically signed. This communication has been approved according to ECHA's internal decision-approval process.

**Appendix A: Reasons to request information required under Annex VII of REACH****1. Water solubility**

Water solubility is an information requirement under Annex VII to REACH (Section 7.7).

You have provided the following information:

- i. Study similar to OECD TG 105, ■ (2007).

We have assessed this information and identified the following issue:

To fulfil the information requirement, a study must comply with the OECD TG 105 or the EU Method A.6 (Article 13(3) of REACH). Therefore, the following specifications must be met:

- the shake-flask method is applicable to test material with a water solubility  $\geq 10$  mg/L;
- solids are pulverized before testing;
- the test is conducted with a loading of about five times the quantity required to saturate a given volume of water;
- three flasks are included which are shaken/stirred for 24, 48 and 72 hours, respectively;
- after shaking/stirring, each flask is equilibrated for 24 hours at 20°C;
- the results are considered acceptable, if the results of the flasks shaken for 48 and 72 hours differ by  $\leq 15\%$ . If the results shows a tendency of higher solubility with longer shaking/stirring period, the test is repeated with longer equilibration times;
- a reliable analytical method is available.

Your registration dossier provides a study showing the following:

- the water solubility was determined to be 3.3  $\mu\text{g/L}$ , hence below 10 mg/L;
- the fact that the test material was pulverized or not before testing is not reported;
- about 5 mg of the test sample were suspended in 30 mL water in a sample flask;
- triplicate test samples were shaken for two hours at 30°C (+/- 2°C) and then at ambient temperature (c.a. 22-23°C) for 70 hours;
- the test material concentration was determined UV-VIS. The calibration curve was produced using chloroform as solvent. The lowest calibration point corresponded to a nominal concentration of 0.70 mg/L (absorbance at 514 nm of 0.0555 measured in chloroform with a 10 mm cuvette). The measured absorbance for the test material ranged from 0.00171 to 0.00293 (measured in water with a 100 mm cuvette).

Based on the above, the shake-flask method described in OECD TG 105 is not applicable to the Substance as its solubility is estimated to be well below 10 mg/L. Furthermore, the test design, the loading rate and the sample preparation method are not compliant with the guideline requirements. Finally, the analytical method used in this study did not allow providing a reliable estimate of dissolved concentration. The measured absorbance values in the test samples are more than an order of magnitude below the absorbance value of the lowest calibration point. Considering the inherent uncertainty related to the measurement of low absorbance values and the fact that the calibration curve and test samples use different solvents (i.e. chloroform versus water, which have different  $\lambda_{\text{max}}$ ), the reliability of the reported analytical method is not demonstrated.

On the basis of the above, the information requirement is not fulfilled.

*Study design*

Considering the properties of the Substance (solubility < 10 mg/L), the column elution described in EU A.6/OECD TG 105 is the most appropriate method to fulfil the information requirement for the Substance.

## 2. Partition coefficient n-octanol/water

Partition coefficient in n-octanol/water is an information requirement under Annex VII to REACH (Section 7.8).

You have provided the following information:

- Study similar to OECD TG 107, ■ (2007).

We have assessed this information and identified the following issues:

- A. To fulfil the information requirement, a study must comply with the OECD TG 107 or OECD TG 117 or OECD TG 123 or the EU Method A.8 (Article 13(3) of REACH). These test guidelines describe three methods (the shake flask method, the HPLC method and the slow-stirring method) for conducting the determining the partition coefficient between water and n-octanol (Log Kow). The EU test method A.8 specifies that the method selection must be based on the properties of the substance and on a preliminary determination of Log Kow using the individual solubilities of the test material in water and n-octanol. This preliminary estimate is considered sufficient only if none of the recommended method are technically feasible due to specific substance properties (e.g. surface active substances).

Your registration dossier provides a study claimed similar to OECD TG 107. However, the robust study summary reports that the study was conducted according to the ETAD method where log Kow is determined using the individual solubilities of the test material in water and n-octanol. You have not provided any justification as to why none of the methods listed above are technically feasible.

- B. To provide an acceptable determination of the partition coefficient using individual solubilities in water and n-octanol, the calculation must be based on reliable individual solubilities estimates.

You used the information discussed under Section A.1 as the water solubility estimated used in the calculation. You report that the n-octanol solubility estimate was determined using a similar method.

As explained under Section A.1, the information provided in your registration does not fulfil the information requirement. Furthermore, as a similar approach was used to determine n-octanol solubility, similar issues identified under Section A.1 also apply to the determination of n-octanol solubility. Hence, the log Kow value reported in your registration dossier is not reliable.

On the basis of the above, the information requirement is not fulfilled.

### *Study design*

Considering the properties of the Substance (sparingly soluble particles), the Partition Coefficient (n-octanol/water), HPLC Method (test method: OECD TG 117) or alternatively the Partition Coefficient (1-Octanol/Water): Slow-Stirring Method (test method: OECD TG 123) are the most appropriate method to fulfil the information requirement for the Substance.

### **3. In vitro cytogenicity study in mammalian cells or In vitro micronucleus study**

Under Annex VII, Section 8.4, column 2 of REACH, further mutagenicity studies must be considered in case of a positive result in an *in vitro* gene mutation study in bacteria. The ECHA guidance R.7a<sup>4</sup> further specifies that "*REACH Annex VII substances for which only a bacterial gene mutation test has been conducted and for which the result is positive should be studied further, according to the requirements of Annex VIII.*" This is for the reason that the *in vitro* cytogenicity test under Section 8.4.2 will allow to further investigate the mutagenicity of the substance in accordance with the REACH integrated testing strategy. The obtained *in vitro* data will inform on the genotoxic concern(s) associated with the substance and help identify the most adequate follow-up *in vivo* study (same *in vivo* study requested under A.5. and B.2).

For the assessment, selection and specifications of the study to be performed, see section B.1.

### **4. In vivo genetic toxicity study**

Under Annex VII, Section 8.4, column 2 of REACH, further mutagenicity studies must be considered in case of a positive result in an *in vitro* gene mutation study in bacteria.

The ECHA guidance R.7a<sup>5</sup> states that following a positive result in an *in vitro* test, "*adequately conducted somatic cell in vivo testing is required to ascertain if this potential can be expressed in vivo. In cases where it can be sufficiently deduced that a positive in vitro finding is not relevant for in vivo situations (e.g. due to the effect of the test substances on pH or cell viability, in vitro-specific metabolism: see also Section R.7.7.4.1), or where a clear threshold mechanism coming into play only at high concentrations that will not be reached in vivo has been identified (e.g. damage to non-DNA targets at high concentrations), in vivo testing will not be necessary.*"

Your dossier contains positive results for the *in vitro* gene mutation study in bacteria which raise the concern for gene mutation.

ECHA considers that an appropriate *in vivo* follow up genetic toxicity study is necessary to address the concern identified *in vitro*.

For the assessment, selection and specifications of the study to be performed, see section B.2.

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<sup>4</sup> ECHA Guidance R.7a, section R.7.7.6.3, p.570.

<sup>5</sup> ECHA Guidance R.7a, section R.7.7.6.3, p.570.

**Appendix B: Reasons to request information required under Annex VIII of REACH****1. In vitro cytogenicity study in mammalian cells or In vitro micronucleus study**

An *in vitro* cytogenicity study in mammalian cells or an *in vitro* micronucleus study is an information requirement under Annex VIII to REACH (Section 8.4.2).

You have provided the following information:

- i. *In vitro* chromosome aberration study in mammalian cells with the Substance (██████████, 1992), according to OECD TG 473.

You also provided an *in vivo* study in your dossier:

- ii. Unscheduled DNA Synthesis (UDS) Test with Mammalian Liver Cells *in vivo* with the Substance (██████████, 2013), according to OECD TG 486.

Although you do not explicitly claim an adaptation, ECHA notes that the studies i. are not performed according to Good Laboratory Practice (GLP) as required under Article 13(4). We understand that you may have submitted these studies as an adaptation according to Annex XI, Section 1.1.2.

With the submission of the *in vivo* studies ii., we understand that you have attempted to adapt this information requirement under Section 8.4.2., Column 2, first indent, Annex VIII to REACH.

We have assessed this information and identified the following issues:

**A. Non GLP study and adaptation under section 1.1.2 of Annex XI**

An adaptation according to Annex XI, Section 1.1.2 enables registrants to claim that the data from experiments not carried out according to GLP or the test methods referred to in Article 13(3) can be considered equivalent to data generated by those test methods.

The adaptation rule in Annex XI, Section 1.1.2. imposes a number of cumulative conditions for an adaptation to be valid, in particular:

1. Adequate and reliable coverage of the key parameters foreseen to be investigated in the corresponding test method (OECD TG 473/ 487), which include among other:
  - a) three experimental conditions: a short treatment with and without metabolic activation and a long treatment without metabolic activation
  - b) cells should be exposed to the test chemical with and without metabolic activation for 3-6 hours
  - c) data on the cytotoxicity and solubility limitations must be reported.
2. Adequate and reliable documentation of the study is provided. More specifically, Article 10(a)(vii) and Article 3(28) require documentation studies to be reported in the form of a robust study summary.

The reported data for the study i. you have provided did not include:

- a) the long treatment without the metabolic activation
- b) cell exposure to test chemical with metabolic activation for 3-6 hours
- c) data on the cytotoxicity and solubility limitations.

Therefore, the study i. is not adequate and an adaptation under section 1.1.2. cannot be accepted.

**B. Adaptation under column 2 of Section 8.4.2 of Annex VIII to REACH**

Under Section 8.4.2., Column 2, first indent, Annex VIII to REACH, the study may be omitted if adequate data from an *in vivo* cytogenicity test are available. The *in vivo* study must be either a micronucleus test or a chromosomal aberration test, performed according to OECD TG 474 or 475, respectively<sup>6</sup>.

However, you have provided an Unscheduled DNA Synthesis (UDS) Test with Mammalian Liver Cells *in vivo* (OECD TG 486)

This test is not a micronucleus test or a chromosomal aberration test. Therefore, the requirements of Section 8.4.2., Column 2, first indent, Annex VIII to REACH are not met.

On the basis of the above, the information requirement is not fulfilled.

*Study design*

To fulfil the information requirement for the Substance, either *in vitro* cytogenicity study in mammalian cells (test method OECD TG 473) or *in vitro* micronucleus study (test method OECD TG 487) are considered suitable.

**2. In vivo genetic toxicity study**

Under Annex VIII, Section 8.4, column 2 of REACH, the performance of an appropriate *in vivo* somatic cell genotoxicity study must be considered if there is a positive result in any of the *in vitro* genotoxicity studies in Annex VII or VIII.

Your dossier contains positive results for the *in vitro* gene mutation study in bacteria which raise the concern for gene mutation.

You have provided the following information:

- Unscheduled DNA Synthesis (UDS) Test with Mammalian Liver Cells *in vivo* with the Substance (██████, 2013), according to OECD TG 486.

We have assessed this information and identified the following issue:

According to the ECHA Guidance Chapter R.7a<sup>7</sup>, the transgenic rodent somatic and germ cell gene mutation assay ("TGR assay", OECD TG 488) and the *in vivo* mammalian alkaline comet assay ("comet assay", OECD TG 489) are suitable to follow up a *positive in vitro* result on gene mutation. Therefore to be considered adequate, the study has to meet the requirements of OECD TG 489 or 488.

However, you provided a study according to OECD TG 486.

This test is neither a transgenic rodent somatic and germ cell mutation assay nor a comet assay.

Therefore, the provided *in vivo* test is not adequate.

On the basis of the above, an appropriate *in vivo* follow-up mutagenicity study is necessary to address the concern identified *in vitro*.

<sup>6</sup> ECHA Guidance R.7a, Table R.7.7-3, p.558

<sup>7</sup> ECHA Guidance Chapter R.7a, Section R.7.7.6.3



*i. Test selection*

As indicated above, the TGR and the comet assay are suitable tests to follow up the concern on gene mutation for the Substance.

However, this decision also requests an *in vitro* test (see Section B.1), which could raise a concern for chromosomal aberration in case of positive results.

In case there is also a concern for chromosomal aberration, you must combine the comet assay and the *in vivo* mammalian erythrocyte micronucleus test ("MN test", OECD TG 474) into a single study. The MN test is a mutagenicity test that provides evidence on *in vivo* chromosomal mutagenicity, as the study detects both structural and numerical chromosomal aberrations. The combined study can help reduce the number of tests performed and the number of animals used while addressing both chromosomal aberration and gene mutation.

Therefore, you must wait for the result of the *in vitro* test requested under B.1 and, depending on the result, to conduct either a) Comet assay or TGR, if the test results of request B.1 are negative; or b) Comet assay combined with MN test if the test results of request B.1 are positive. The deadline set in this decision allows for sequential testing.

*ii. Study design**a) Comet assay or TGR assay (if the test results of request B.1 are **negative**)**Comet assay:*

According to the test method OECD TG 489, the test must be performed in rats. Having considered the anticipated routes of human exposure and adequate exposure of the target tissue(s) performance of the test by the oral route is appropriate.

In line with the test method OECD TG 489, the test must be performed by analysing tissues from liver as primary site of xenobiotic metabolism, glandular stomach and duodenum as sites of contact. There are several expected or possible variables between the glandular stomach and the duodenum (different tissue structure and function, different pH conditions, variable physico-chemical properties and fate of the Substance, and probable different local absorption rates of the Substance and its possible breakdown product(s)). In light of these expected or possible variables, it is necessary to analyse both tissues to ensure a sufficient evaluation of the potential for genotoxicity at the site of contact in the gastro-intestinal tract.

*TGR assay:*

According to the test method OECD TG 488, the test must be performed in transgenic mice or rats and the test substance is usually administered orally.

Based on the recent update<sup>8</sup> of OECD TG 488, you are requested to follow the new 28+28d regimen, as it permits the testing of mutations in somatic tissues and as well as in tubule germ cells from the same animals. This updated version provides for a transitional period for the new version. However, ECHA is aware that testing according to the updated OECD TG is already available from CROs and the new study design would provide meaningful germ cell data, so this decision requires the application of the new version.

According to the test method OECD TG 488, the test must be performed by analysing tissues from liver as slowly proliferating tissue and primary site of xenobiotic metabolism, glandular

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<sup>8</sup> The updated OECD TG 488, adopted on 26 June 2020, is available on OECD website at <https://www.oecd-ilibrary.org/docserver/9789264203907-en.pdf?expires=1596539942&id=id&accname=guest&checksum=D552783C4CB0FC8045D04C88EFFBFA66>

stomach and duodenum as rapidly proliferating tissue and site of direct contact. There are several expected or possible variables between the glandular stomach and the duodenum (different tissue structure and function, different pH conditions, variable physico-chemical properties and fate of the Substance, and probable different local absorption rates of the Substance and its possible breakdown product(s)). In light of these expected or possible variables, it is necessary to analyse both tissues to ensure a sufficient evaluation of the potential for mutagenicity at the site of contact in the gastro-intestinal tract. However, duodenum must be stored (at or below  $-70\text{ }^{\circ}\text{C}$ ) until the analysis of liver and glandular stomach is completed; the duodenum must then be analysed only if the results obtained for the glandular stomach and for the liver are negative or inconclusive.

#### *Germ cells*

In case you decide to perform the comet assay, you may consider to collect the male gonadal cells from the seminiferous tubules in addition to the other aforementioned tissues in the comet assay, as it would optimise the use of animals. You can prepare the slides for male gonadal cells and store them for up to 2 months, at room temperature, in dry conditions and protected from light. Following the generation and analysis of data on somatic cells in the comet assay, you should consider analysing the slides prepared with gonadal cells.

In case you decide to perform the TGR, you may consider to collect the male germ cells (from the seminiferous tubules) at the same time as the other tissues, in order to limit additional animal testing. According to the OECD 488, the tissues (or tissue homogenates) can be stored under specific conditions and used for DNA isolation for up to 5 years (at or below  $-70\text{ }^{\circ}\text{C}$ ). This duration is sufficient to allow you or ECHA, to decide on the need for assessment of mutation frequency in the collected germ cells.

This type of evidence may be relevant for the overall assessment of possible germ cell mutagenicity including classification and labelling according to the CLP Regulation.

#### *b) Comet assay combined with MN test (if the test results of request B.1 are **positive**)*

According to the test method OECD TG 489, the test must be performed in rats. Therefore, the combined test (OECD TG 489 and OECD TG 474) must be performed in rats. Having considered the anticipated routes of human exposure and the need for adequate exposure of the target tissue(s) performance of the test by the oral route is appropriate.

In line with the test method OECD TG 489, the test must be performed by analysing tissues from liver as primary site of xenobiotic metabolism, glandular stomach and duodenum as sites of contact. There are several expected or possible variables between the glandular stomach and the duodenum (different tissue structure and function, different pH conditions, variable physico-chemical properties and fate of the Substance, and probable different local absorption rates of the Substance and its possible breakdown product(s)). In light of these expected or possible variables, it is necessary to analyse both tissues to ensure a sufficient evaluation of the potential for genotoxicity at the site of contact in the gastro-intestinal tract.

The combination of OECD TGs 489 and 474 should not impair the validity of and the results from each individual study. Careful consideration should be given to the dosing, and tissue sampling for the comet analysis alongside the requirements of tissue sampling for the mammalian erythrocyte micronucleus test (see OECD TG 489, e.g. Bowen *et al.* 2011<sup>9</sup>).

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<sup>9</sup> Bowen D.E. et al. 2011. Evaluation of a multi-endpoint assay in rats, combining the bone-marrow micronucleus test, the comet assay and the flow-cytometric peripheral blood micronucleus test. *Mutation Research* 722 7–19

*Germ cells*

You may consider to collect the male gonadal cells from the seminiferous tubules in addition to the other aforementioned tissues in the comet assay, as it would optimise the use of animals. You can prepare the slides for male gonadal cells and store them for up to 2 months, at room temperature, in dry conditions and protected from light. Following the generation and analysis of data on somatic cells in the comet assay, you should consider analysing the slides prepared with gonadal cells. This type of evidence may be relevant for the overall assessment of possible germ cell mutagenicity including classification and labelling according to the CLP Regulation.

## **Appendix C: Requirements to fulfil when conducting and reporting new tests for REACH purposes**

### **A. Test methods, GLP requirements and reporting**

1. Under Article 13(3) of REACH, all new data generated as a result of this decision must be conducted according to the test methods laid down in a European Commission Regulation or to international test methods recognised by the Commission or ECHA as being appropriate.
2. Under Article 13(4) of REACH, ecotoxicological and toxicological tests and analyses must be carried out according to the GLP principles (Directive 2004/10/EC) or other international standards recognised by the Commission or ECHA.
3. Under Article 10(a)(vi) and (vii) of REACH, all new data generated as a result of this decision must be reported as study summaries, or as robust study summaries, if required under Annex I of REACH. See ECHA Practical Guide on How to report robust study summaries<sup>10</sup>.

### **B. Test material**

Before generating new data, you must agree within the joint submission on the chemical composition of the material to be tested (Test Material) which must be relevant for all the registrants of the Substance.

1. Selection of the Test material(s)

The Test Material used to generate the new data must be selected taking into account the following:

- the variation in compositions reported by all members of the joint submission,
  - the boundary composition(s) of the Substance,
  - the impact of each constituent/ impurity on the test results for the endpoint to be assessed. For example, if a constituent/ impurity of the Substance is known to have an impact on (eco)toxicity, the selected Test Material must contain that constituent/ impurity.
2. Information on the Test Material needed in the updated dossier
    - You must report the composition of the Test Material selected for each study, under the "Test material information" section, for each respective endpoint study record in IUCLID.
    - The reported composition must include all constituents of each Test Material and their concentration values and other parameters relevant for the property to be tested.

This information is needed to assess whether the Test Material is relevant for the Substance and whether it is suitable for use by all members of the joint submission.

Technical instructions on how to report the above is available in the manual on How to prepare registration and PPORD dossiers<sup>11</sup>.

<sup>10</sup> <https://echa.europa.eu/practical-guides>

<sup>11</sup> <https://echa.europa.eu/manuals>

## **Appendix D: Procedure**

The Substance is listed in the Community rolling action plan (CoRAP) for the start of substance evaluation in 2019/2020.

This decision does not prevent ECHA from initiating further compliance checks at a later stage on the registrations present.

ECHA followed the procedure detailed in Articles 50 and 51 of REACH.

The compliance check was initiated on 24 March 2020.

ECHA notified you of the draft decision and invited you to provide comments.

Comments related to registration issues were provided by one addressee. These were addressed through a separate communication.

ECHA did not receive any comments within the commenting period on the requests listed in the decision.

ECHA notified the draft decision to the competent authorities of the Member States for proposals for amendment.

ECHA received proposal(s) for amendment and modified the draft decision.

ECHA invited you to comment on the proposed amendment(s) and referred the modified draft decision to the Member State Committee.

You did not provide any comments on the proposed amendment(s).

The Member State Committee reached a unanimous agreement on the draft decision during its MSC-74 meeting and ECHA took the decision according to Article 51(6) of the REACH Regulation.

**Appendix E: List of references - ECHA Guidance<sup>12</sup> and other supporting documents**Evaluation of available information

Guidance on information requirements and chemical safety assessment, Chapter R.4 (version 1.1., December 2011), referred to as ECHA Guidance R.4 where relevant.

QSARs, read-across and grouping

Guidance on information requirements and chemical safety assessment, Chapter R.6 (version 1.0, May 2008), referred to as ECHA Guidance R.6 where relevant.

Read-across assessment framework (RAAF, March 2017)<sup>13</sup>

RAAF - considerations on multiconstituent substances and UVCBs (RAAF UVCB, March 2017)<sup>13</sup>

Physical-chemical properties

Guidance on information requirements and chemical safety assessment, Chapter R.7a (version 6.0, July 2017), referred to as ECHA Guidance R.7a in this decision.

Toxicology

Guidance on information requirements and chemical safety assessment, Chapter R.7a (version 6.0, July 2017), referred to as ECHA Guidance R.7a in this decision.

Guidance on information requirements and chemical safety assessment, Chapter R.7c (version 3.0, June 2017), referred to as ECHA Guidance R.7c in this decision.

Environmental toxicology and fate

Guidance on information requirements and chemical safety assessment, Chapter R.7a (version 6.0, July 2017), referred to as ECHA Guidance R.7a in this decision.

Guidance on information requirements and chemical safety assessment, Chapter R.7b (version 4.0, June 2017), referred to as ECHA Guidance R.7b in this decision.

Guidance on information requirements and chemical safety assessment, Chapter R.7c (version 3.0, June 2017), referred to as ECHA Guidance R.7c in this decision.

PBT assessment

Guidance on information requirements and chemical safety assessment, Chapter R.11 (version 3.0, June 2017), referred to as ECHA Guidance R.11 in this decision.

Guidance on information requirements and chemical safety assessment, Chapter R.16 (version 3.0, February 2016), referred to as ECHA Guidance R.16 in this decision.

Data sharing

Guidance on data-sharing (version 3.1, January 2017), referred to as ECHA Guidance on data sharing in this decision.

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<sup>12</sup> <https://echa.europa.eu/guidance-documents/guidance-on-information-requirements-and-chemical-safety-assessment>

<sup>13</sup> <https://echa.europa.eu/support/registration/how-to-avoid-unnecessary-testing-on-animals/grouping-of-substances-and-read-across>

OECD Guidance documents<sup>14</sup>

Guidance Document on aqueous-phase aquatic toxicity testing of difficult test chemicals – No 23, referred to as OECD GD 23.

Guidance document on transformation/dissolution of metals and metal compounds in aqueous media – No 29, referred to as OECD GD 29.

Guidance Document on Standardised Test Guidelines for Evaluating Chemicals for Endocrine Disruption – No 150, referred to as OECD GD 150.

Guidance Document supporting OECD test guideline 443 on the extended one-generation reproductive toxicity test – No 151, referred to as OECD GD 151.

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<sup>14</sup> <http://www.oecd.org/chemicalsafety/testing/series-testing-assessment-publications-number.htm>

**Appendix F: Addressees of this decision and the corresponding information requirements applicable to them**

You must provide the information requested in this decision for all REACH Annexes applicable to you.

<b>Registrant Name</b>	<b>Registration number</b>	<b>Highest REACH Annex applicable to you</b>
[REDACTED]	[REDACTED]	[REDACTED]
[REDACTED]	[REDACTED]	[REDACTED]
[REDACTED]	[REDACTED]	[REDACTED]
[REDACTED]	[REDACTED]	[REDACTED]
[REDACTED]	[REDACTED]	[REDACTED]

Where applicable, the name of a third party representative (TPR) may be displayed in the list of recipients whereas ECHA will send the decision to the actual registrant.